



## Chemopreventive effect of thymoquinone on benzo(a)pyrene-induced lung cancer in male swiss albino mice

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### ABSTRACT

Benzo[a]pyrene [B(a)P], a well-known environmental carcinogen, promotes oxidative stress and DNA damage. Thymoquinone (TQ), the main active constituent of black seed essential oil, exhibits promising effects against inflammatory diseases and cancer. The present study was designed to investigate the possible protective effect of TQ on [B(a)P]-induced lung cancer in mice. One hundred male Swiss Albino mice were divided into four equal groups. Group I :( Control group) received no drugs. Group II :( lung cancer- induced group) mice administered with a single dose of [B(a)P] (100 mg/ kg b.wt, intraperitoneally). Group III :( lung cancer + TQ treated group) mice injected with [B(a)P] as in group II and treated with TQ (20 mg/kg b.wt/day, orally) from 22th week to 30th weeks. Group IV: (lung cancer + TQ protected group) mice received TQ (20 mg/kg b.wt. / Orally) on alternate days from 1 day prior to [B(a)P] injection and were treated continuously with TQ until 30th week (end of experiment). Blood samples and lung tissue specimens were collected at the end of experimental period (30 week) for determination of serum carcino-embryonic antigen (CEA), Haptoglobin (HPT) and Gamma glutamyl transferase ( $\gamma$ -GT) in addition to catalase (CAT), Super oxide dismutase(SOD), L-Malondialdehyde (L-MDA), Caspase3, DNA fragmentation(DF), Cyclooxygenase -2(COX-2) in lung tissues. The obtained results revealed that, [B(a)P] potentially increased serum  $\gamma$ -GT activity, HP and CEA levels in addition to lung tissues COX-2, Caspase 3 gene, L-MDA and DNA fragmentation. However, SOD and GST activities in lung tissues were significantly decreased. TQ treatment was able to mitigate lung cancer induced by [B(a)P] through enhanced the activity of SOD, CAT and attenuated the increased caspase 3 gene, DNA fragmentation, COX-2 and L-MDA in lung tissues and serum CEA, HP and  $\gamma$ -GT. It could be concluded that, TQ may be effective in reducing lung cancer by its radical scavenging activity and anti-inflammatory effect, regenerating endogenous antioxidant mechanisms and decreased caspase-3 gene and DNA fragmentation in lung tissues. These results suggest that, the possible efficiency of TQ as an distinct chemo-preventive agent in lung carcinogenesis.

**KEYWORDS:** thymoquinone, Benzo(a)pyrene, Lung cancer, DNA fragmentation, caspase-3 gene, Cyclooxygenase -2.

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### 1. INTRODUCTION

Lung cancer is one of the most lethal cancers of the 20th century and still the most common cancer in the world causing up to 3 million deaths annually, and it is increasing at a rapid rate (Hecht et al., 2002; Osann et al., 2000). In Egypt, official statistics showed that lung cancer is the second most common cancer in men and second leading cause of cancer death, after

bladder cancer. Mice lung tumor-genesis systems be valuable tools to study the process of chemical carcinogenesis induced by polycyclic aromatic hydrocarbons (PAHs). PAHs and *N*-nitrosamines are the two major classes of tobacco-related inhaled carcinogens (Kamaraj et al., 2007). Benzo(a)pyrene [B(a)P] is the archetypal PAH as it is the most intensely studied

PAH, it is ubiquitous in the environment and it is a very potent carcinogen (Cavaliere *et al.*, 1991). Who added that, [B(a)P] is typically selected as the standard against which the cancer potency of other PAHs are tested. Furthermore, PAH including [B(a)P], a potent tobacco carcinogen. Moreover, [B(a)P] induces cancer in many species of rodents and [B(a)P] itself as well as [B(a)P]-containing complex environmental mixtures are known human respiratory carcinogens (Straif, 2005 and IARC, 1989).

Previous studies have proved that the toxicity of [B(a)P] behind its intermediate metabolites and the oxidative damage caused by reactive oxygen species (ROS) (Wang *et al.*, 2009; Wester *et al.*, 2012). Additionally, PAH is a significant pro-carcinogenic substance, which requires metabolic activation to electrophilic reactive metabolites for its carcinogenic activity (Gelboin *et al.*, 1980).

Moreover, DNA damage has been recognized as the onset of many diseases, including cancer and could be a useful biomarker of the oxidative status and antioxidant defense system of an organism. On the other hand, smoking is undoubtedly the main risk factor, to which 90% of lung cancer cases are attributable (Hecht, 1999; Ruano-Ravina *et al.*, 2003 and Winterhalder *et al.*, 2004). In fact, (ROS) and organic free radical intermediates formed from many carcinogens are suggested to be involved in the initiation and progression of carcinogenic transformation (Panandiker *et al.*, 1994).

Cancer chemoprevention can be defined as the prevention, inhibition or reversal of carcinogenesis by administration of one or more chemical entities, either as individual drugs or as naturally occurring constituents of the diet (Hanspeter *et al.*, 2000; Anto *et al.*, 2002).

Numerous studies have shown that the seeds and oil of this plant are characterized by a very low degree of toxicity (Ali and

Blunden, 2003). Furthermore, TQ has been shown to exert anti-inflammatory, anti-oxidant and anti-neoplastic effects both in vitro and in vivo (Pagola *et al.*, 2004). Many investigators have shown that the growth inhibitory effects of TQ are specific to cancer cells (Gali-Muhtasib *et al.*, 2004; Shoieb *et al.*, 2003; Worthen *et al.*, 1998). In addition TQ also exerts anti-oxidant effects and inhibits inflammation in animal models and cell culture systems (Mansour *et al.*, 2002). Accordingly, the present study was designed to evaluate the chemopreventive activity and the potential protective effect of TQ against [B(a)P] induced lung carcinogenesis in Swiss albino mice by determination of antioxidant parameters like catalase (CAT), Super oxide dismutase (SOD), L-Malondialdehyde (L-MDA), Caspase 3, DNA fragmentation and Cyclooxygenase-2 (COX-2) in lung tissues in addition to some serum parameters as carcino embryonic antigen (CEA), Haptoglobin (HP) and Gamma glutamyl transferase ( $\gamma$  GT).

## 2. MATERIAL AND METHODS

### 2.1. Experimental animals:

One hundred male Swiss Albino mice of 6-8 weeks old and weighing 25-30 gm were used in the experimental investigation of this study. Mice were obtained from the Laboratory Animals Research Center, Faculty of Veterinary Medicine, Benha University. The animals were housed in separated metal cages and kept at constant environmental and nutritional conditions throughout the period of the experiment. Fresh and clean drinking water was supplied ad-libitum. The animals were left for 15 days for acclimatization prior to the beginning of the experiment.

#### 2.1.1. Thymoquinone:

It is [2-isopropyl-5-methyl-1, 4 benzoquinone]. Thymoquinone yellow crystals, not soluble in water but dissolved

in organic solvents (as ethyl alcohol). TQ have been purchased by Sigma Chemical Co. (St. Louis, Mo, USA) and purchased from Schnellendorf, Germany through the Egyptian International Center for Import Cairo, Egypt.

### 2.1.2. Preparation and Dosage of Thymoquinone :

Thymoquinone was dissolved in few drops of ethyl alcohol and complete with distilled water and administered at dosage of (20 mg/kg.b.wt./ day, orally) all over the experimental period (Gali-Muhtasib et al., 2006)

### 2.1.3. Induction of lung cancer :

[B(a)P] was freshly dissolved in corn oil to ensure the stability of the chemical just prior to use. Lung cancer was induced in mice by a single intraperitoneal injection of [B (a) P] at a dose of (100 mg/kg body weight) (Magesh et al., 2009). [B (a) P] has been purchased by Sigma Chemical Co. (St. Louis, Mo, USA) and purchased from Schnellendorf, Germany through the Egyptian International Center for Import Cairo, Egypt.

### 2.2. Experimental design :

Mice were randomly divided into four main equal groups, 25 animal each, placed in individual cages and classified as follow:

Group (1): Control Normal Group:

Mice received no drugs, served as untreated control for all experimental groups.

Group II :( lung cancer- induced group:(

Mice administered with a single dose of [B (a) P] (100 mg/ kg b.wt, intraperitoneally), served as carcinogenic non treated group.

Group III :( lung cancer + TQ treated group:(

Mice injected with [B(a)P] (100 mg/ kg b.wt, intraperitoneally) and treated with TQ (20 mg/kg b.wt/day, Orally) from 22th week of the experiment and continued to 30th weeks(end of the experiment ).

Group IV :( lung cancer + TQ protected group:(

Mice received TQ (20 mg/kg b.wt. / Orally) on alternate days from 1 day prior to [B(a)P] injection and were treated continuously with TQ until 30th week (end of experiment).

### 2.3. Sampling :

Blood samples and tissue specimens (lung tissues) were collected at the end of the experiment on 30th weeks from all animal groups (control and experimental group)

#### 2.3.1. Blood samples:

Blood samples for serum separation were collected by ocular vein puncture at the end of each experimental period in dry, clean, and screw capped tubes and serum were separated by centrifugation at 2500 r.p.m for 15 minutes. The clean, clear serum was separated by automatic pipette and received in dry sterile samples tube and kept in a deep freeze at -20 °C until used for subsequent biochemical analysis. All sera were analyzed for determination of (HPT), (α GT) and (CEA)

#### 2.3.2. Tissue samples (lung tissue)

At the end of the experimental period, the animals were sacrificed by cervical decapitation. The lungs were dissected out, quickly removed and were rinsed in ice-cold physiological saline, then blotted between 2 filter papers and quickly stored in a deep freezer at -20 °C for further biochemical analysis. Briefly, lung tissue was subsequently minced into small pieces and 10% homogenate was prepared in cold phosphate buffer (pH 7.4). The homogenate

was centrifuged at  $1000 \times g$  for 10 min at  $4^{\circ}\text{C}$ , and the supernatant was used directly for the determination of CAT, SOD, L-MDA, COX-2, Caspase 3 and DNA fragmentation.

#### 2.4. Biochemical analysis:

Serum CEA, HPT,  $\gamma$  GT and lung tissues SOD, CAT, L-MDA, COX-2, Caspase 3 and DNA fragmentation were analyzed.

#### 2.5. Statistical analysis :

The obtained data were statistically analyzed by one-way analysis of variance (ANOVA) followed by the Duncan multiple test. All analyses were performed using the statistical package for social science (SPSS, 13.0 software, 2009). Values of  $P < 0.05$  were considered to be significant.

### 3. RESULTS

Protective and treatment Effects of TQ administration on some serum and lung tissues biochemical parameters of [B(a)P]-induced lung cancer in mice. The obtained results demonstrated in (Table 1) revealed that, administration of [B(a)P] induced lung cancer in mice exhibited a significant decrease in CAT and SOD activities and significantly increased COX-2, Caspase 3, L-MDA, and DNA fragmentation in lung tissues and in serum HP, CEA levels and  $\gamma$ GT activity when compared with normal control group. protection and treatment with TQ in [B(a)P] induced lung cancer in mice significantly increased CAT and SOD activities and significantly decreased and attenuated the increased in COX-2, Caspase 3, L-MDA, and DNA fragmentation in lung tissues. Also, TQ administration significantly reduced elevated serum  $\gamma$  GT activity, HP and CEA concentrations when compared with [B(a)P]-induced lung cancer non-treated group.

### 4. DISCUSSION

Lung cancer is currently a leading cause of death all over the world. In recent years, considerable attention has been given to increased dietary intake of phytochemicals, since numerous epidemiological as well as experimental studies gave positive correlation between reduced risk of cancer and intake of phytochemicals (Ramakrishnan *et al.*, 2007). Experimental studies have discovered that the process of carcinogenesis can be modulated. One of the approaches is chemoprevention by administering or consuming foods and drinks containing chemo-preventive agents (Ren *et al.*, 2003). The present study clearly demonstrates a potent inhibitory activity of TQ, the main constituent of the volatile oil of *Nigella sativa* (Black seed) against [B(a)P]-induced mutagenic effect in lung tissue in male mice. The use of cytotoxic agents plays an important role in the management of intermediate and high-risk tumors in addition to delayed surgery. Numerous studies have shown that the seeds and oil of this plant are characterized by a very low degree of toxicity (Ali and Blunden, 2003). In the present study, [B(a)P] treated mice showed a significant increase in the serum  $\gamma$  GT activity and CEA and HPT concentrations when compared with control group. However, TQ administration significantly reduced elevated serum  $\gamma$  GT activity, HPT and CEA concentrations when compared with [B(a)P]-induced lung cancer non-treated group. Similarly, a significant increase in the expression of serum CEA and serum marker enzyme  $\gamma$  GT activity were observed in the [B(a)P] administered group. Also, (Kassie *et al.*, 2007) observed that, HPT (2004). These toxic radicals are involved in mediating tissue lipid peroxidation. Lipid peroxidation-induced tissue damage is the sensitive feature in the cancerous conditions any deterioration or destruction of the membrane can lead to the leakage of these enzymes from the tissues (Ramakrishnan *et*

Table 1: Protective and treatment Effects of TQ administration on some serum and lung tissues biochemical parameters of B(a)P-induced lung cancer in mice at the end o experimental period (30 days).

Experimental groups	Control Normal group	B(a)P group	B(a)P + TQ treated group	B(a)P + TQ protected group
Parameters				
CEA (ng/ml)	0.29±0.08 <sup>b</sup>	1.59±0.27 <sup>a</sup>	0.58±0.10 <sup>b</sup>	0.44±0.13 <sup>b</sup>
Haptoglobin(ng/ml)	2.05±0.21 <sup>b</sup>	7.40±1.31 <sup>a</sup>	2.67±0.37 <sup>b</sup>	2.17±0.60 <sup>b</sup>
GGT (U/L)	25.60±6.11 <sup>b</sup>	125.73±15.87 <sup>a</sup>	47.40±8.89 <sup>b</sup>	28.40±7.00 <sup>b</sup>
COX-2 (U/g.tissue)	4.88±0.48 <sup>c,d</sup>	12.42±1.04 <sup>a</sup>	5.62±0.41 <sup>b,c</sup>	3.05±0.84 <sup>d</sup>
Caspase-3 gene activity	0.59±0.05 <sup>c</sup>	2.44±0.03 <sup>a</sup>	1.19±0.19 <sup>b</sup>	0.77±0.11 <sup>c</sup>
SOD (U/g.tissue)	25.70±1.75 <sup>d</sup>	10.19±1.87 <sup>e</sup>	41.19±0.22 <sup>b</sup>	46.58±1.47 <sup>a</sup>
CAT(mmol/g.tissue)	59.04±3.33 <sup>a</sup>	19.06±1.92 <sup>c</sup>	47.75±5.11 <sup>b</sup>	59.26±2.87 <sup>a</sup>
MDA(mmol/g. tissue)	26.40±5.68 <sup>c</sup>	200.86±10.93 <sup>a</sup>	92.74±2.75 <sup>b</sup>	41.27±9.16 <sup>c</sup>
DNA fragmentation %	86.86±25.38 <sup>c</sup>	1477.57±159.42 <sup>a</sup>	144.45±38.71 <sup>c</sup>	115.37±32.69 <sup>c</sup>

Data are presented as (Mean ± S.E). S.E = Standard error. Mean values with different superscript letters in the same column are significantly different at ( $P < 0.05$ ).

level was increased in carcinogenic-treated mice. A very highly significant increase in level of serum CEA was observed in [B(a)P] treated female mice as compared to control group (Shaymaa, 2014). Moreover, (Kamara et al., 2009) reported that, administration of [B(a)P] to mice exhibited significant increase in lung specific tumor marker (CEA) and  $\gamma$  GT. The transfer of  $\gamma$ - glutamyl groups from peptide donors to peptide receptors and aminoacids is the catalytic function of  $\gamma$  GT.  $\gamma$  GT is not only useful in diagnosis but also has prognostic value in malignancies such as lung cancer and malignant melanoma [Obrador et al., 2002]. [B(a)P], a well-identified environmental carcinogen is known to produce enormous amounts of free radicals and these free radicals and non-radical oxidizing species are highly reactive, toxic and mutagenic (Selvendiran et al., and al., 2007). The marked elevation in such serum parameters observed in [B (a) P] treated group may be due to the genotoxic

property of [B(a)P], which is a very effective carcinogen enhancing oxidative stress and consequently inducing free radical formation, which in turn react with lipids in the cell membrane causing lipid peroxidation (Selvendiran et al., 2004). TQ is a well-known scavenger of ROS such as superoxide anions, hydroxyl radicals, and peroxy nitrite anion. In this regard, earlier studies have demonstrated that TQ has a considerable protective effect against reactive oxygen species (ROS) generating agents including significant suppression of fore stomach tumor induced by [B(a)P] (Badary et al., 1999).

In the present study, SOD and CAT activities were significantly decreased and MDA level was significantly increased in lung tissues in [B (a) P] treated mice as compared to normal control mice. Similarly, (Ananda et al., 2013) reported that, [B(a)P] treated mice showed significant increase in the MDA level with

significant decrease of SOD activity in lung tissue as compared to control mice. Also, (Wang *et al.*, 2013) reported that, compared with normal group the value of MDA level, a classic indicator of oxidative stress, was significantly increased in lung tissues of [B(a)P] treated group. Meanwhile, the activity of SOD in [B (a) P] administered group was significantly decreased. Carcinogen induced reactive oxidative species and free radical intermediates have been suggested to have a role in the initiation and development of cancer (Panandiker *et al.*, 1994). [B(a)P] has been reported to cause lipid peroxidation and decrease antioxidant enzymes levels by inducing oxidative stress in lung carcinogenesis (Sikkim and Mulee. 2000). Increased levels of LPO products play a major role in the early phases of tumor growth (Kim *et al.*, 2000). Studies have also shown that SOD activity significantly decrease on [B(a)P] treatment, which may abet in inducing carcinogenesis (Emre *et al.*, 2007). Hence, estimating lipid peroxidation and enzymatic antioxidants like SOD an useful tool for assessing oxidative damage induced carcinogenesis by [B(a)P].

The obtained results revealed that, protection and treatment with TQ in [B(a)P] induced lung cancer in mice significantly increased CAT and SOD activities and attenuated the increased in L-MDA level in lung tissues. This protective effect of TQ may be due to its potential free radical scavenging activity. TQ also exerts anti-oxidant effects and inhibits inflammation in animal models and cell culture systems (Mansour *et al.*, 2002). It is assumed that, these probable anti-apoptogenic effects of TQ may be mediated by one or more of the following mechanisms: Antioxidant activity, immunomodulatory action and genoprotective effects (Rastogi *et al.*, 2010; Gautam *et al.*, 2008; Mousavi *et al.*, 2010; Burits and Bucar; 2000). According to the previous studies, *N. sativa* (TQ source) protects lipids against free-radical damage (Burits and Bucar, 2000). Decreased tissue malondialdehyde(MDA), protein carbonyl levels and prevented inhibition of superoxide dismutase (SOD) and catalase (CAT) enzyme

activities following experimental spinal cord injury in rats were seen following treatment with *N. sativa* (TQ source) (Kanter *et al.*, 2006).

The obtained results revealed that, administration of [B(a)P] induced lung cancer in mice significantly increased COX-2, Caspase 3 and DNA fragmentation in lung tissues when compared with normal control group. This elevation may be due to the genotoxic property of [B(a)P]. The obtained results are nearly similar with those reported by Shaymaa, (2014) She recorded that, [B(a)P] caused significant increase in levels of caspase 3,9 activities in lung tissue compared to control group. Also, Ashish\_ *et al.*, (2008) reported that, [B(a)P] increased the activation of caspase-3,7,8,9 and decreased cell viability. Furthermore, COX-2 has been shown to regulate some aspects of tumor-associated angiogenesis and its expression has been previously reported to be present in elevated levels as compared to normal lung tissue (Anderson *et al.*, 2002). Certain chemo-preventive agents have the capability to affect the COX-2 expression as one of their many functions thereby paving the way for cancer chemoprevention. There are certain agents which have the capability of inhibiting COX-2 and thus have the potential to impart antitumor effects against lung cancer. Phytochemicals such as curcumin and quercetin are such chemopreventive agents which have the potential to affect the COX-2 expression as well as its activity (Aggarwal, 2010).

Preclinical studies do suggest that COX-2 may be involved in the molecular pathogenesis of some types of lung cancer. In lung cancer, COX-2 expression is observed at the majority stages of tumor progression. Clinical studies have demonstrated high levels of expression of COX-2 in almost all non small cell lung cancer pre-invasive precursor lesions as well as invasive lung carcinomas, when compared to normal lung tissue (Anderson *et al.*, 2002). Moreover, increased COX-2 expression is associated with a poor prognosis in lung

cancer (Gardner et al., 2003). Consistent with the preclinical studies, [B(a)P] induced lung carcinogenesis in mice also showed an increase in the activity of COX-2 enzyme in the present study. As inflammation is linked with cancer development and progression (Mascaux et al., 2005), the resulting tumor incidence as well as multiplicity in the lungs of [B(a)P] treated mice can be correlated with the high COX-2 activity.

Similarly, previous work found that, [B(a)P] treatment to mice brought about a statistically significant increase in the activity of COX-2 in the lung tissues of mice. Also, (Zhu et al., 2008) reported that, COX-2 is over expressed in up to 85% of lung cancers and is associated with advanced clinical stage and distant. protection and treatment with TQ in [B(a)P] induced lung cancer in mice attenuated the increased in COX-2, Caspase 3 and DNA fragmentation in lung tissues when compared with [B(a)P] -induced lung cancer non-treated group. Supplementation of phytochemicals significantly reduced elevated activity of COX-2 and further normalized the lung weights as compared to normal controls. Curcumin exhibits its anti-inflammatory effect in part, through inhibition of the NF-kappaB pathway and cyclo-oxygenase 2 (COX-2) enzymes and thus plays a pivotal role in suppressing tumor cell growth (Aggarwal, 2010). On contrary, (Banerjee et al., 2009) indicated that, increased caspase-3 activity was observed in the tumor tissues treated with the TQ.

The present study demonstrated that, TQ administration provided an effective protection and treatment in [B (a) P] induced lung carcinogenesis in Swiss Albino mice.

**In conclusion**, protection and treatment of TQ effectively decrease oxidative stress, ameliorate serum tumor and inflammatory markers, enzymatic antioxidant defense system in lung tissue and protected lung cells via inhibition of caspase 3 and modulating pro-inflammatory enzyme COX-2 activity. This study establishes the role of TQ as a chemo-preventive and chemotherapeutic agent and also, provides the possible

mechanism of TQ modulating caspase 3, DNA fragmentation and inhibiting the activity of COX-2 in [B(a)P] induced lung carcinogenesis.

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### 5. REFERENCES

- Aggarwal, B. B. 2010. Targeting inflammation-induced obesity and metabolic diseases by curcumin and other nutraceuticals, *Annu.Rev. Nutr.*, 21(30):173-99.
- Ali, B. H. and Blunden, G. 2003. Pharmacological and toxicological Properties of *Nigella sativa*. *Phytotherapy Research*, 17, 299–305.
- Ananda, J. R. ,Rijhwania, H. ,Malapatia, K. ,kumara, P., Saikiab,K. and Lakhara, .2013.Anticancer activity of esculetin via – modulation of Bcl-2 and NFB expression in benzo(a)pyrene induced lung carcinogenesis in mice. *biomedicine and preventive nutrition* 3:107-112.
- Anderson, W. F., Umar, A., Viner, J. L. and Hawk, E.T. 2002. The role of cyclooxygenase inhibitors in cancer prevention *Cur. Pharm. Des.* , 8, 1035-62.
- Anto, R. J. Mukhopadyah, A. Denning, K. and Aggarwal, B.B. 2002 Curcumin induces apoptosis through activation of caspase-8, BID cleavage and cytochrome c release: its suppression by ectopic expression of Bcl-2 and Bcl-xl. *Carcino-genesis*; 23:143–50..
- Ashish, Sharma, Aneesh, Neekhra, Ana, L.,Gramajo., Jayaprakash, Patil., Marilyn, Chwa., Baruch, D., Kuppermann, M., Cristina and Kenney. (2008): Effects of Benzo(a)Pyrene, a Toxic Component of Cigarette Smoke, on Human Retinal Pigment Epithelial

- Cells *In vitro*. *Invest. Ophthalmol. Vis. Sci.* 49:11. 5111-5117.
- Badary, O. A., Al-Shabanah, O. A., Nagi, M. N., Al-Rikabi, A. C. and Elmazar, M. M. 1999. Inhibition of benzo(a)pyrene-induced forestomach carcinogenesis in mice by thymoquinone. *European Journal of Cancer Prevention*, 8: 435-440.
- Banerjee, S., Kaseb, A.O., Wang, Z., Kong, D., Mohammad, M. and Padhye, S. 2009. Antitumor activity of gemcitabine and oxaliplatin is augmented by thymoquinone in pancreatic cancer. *Cancer Res*; 69:5575-83.
- Burits, M., and Bucar, F. 2000. Antioxidant activity of *Nigella sativa* essential oil. *Phytother Res*; 14:323-8.
- Cavaliere, E.L., S. Higginbotham, N.V. Rama Krishna, P.D., Devanesan, R. Todorovic, E.G. Rogan, S. and Salmasi. 1991. Comparative dose response tumor-genicity studies of dibenzo [a, l] pyrene versus 7, 12dimethyl-benz[a]anthracene, benzo[a]pyrene and two dibenzo[a,l] pyrenedi hydrodiols in mouse skin and rat mammary gland, *Carcinogenesis* ;12 (10) :1939-1944.
- El Attar, I .2005. Lung cancer in Egypt and neighboring Arab countries: Magnitude of the problem. *UICC*; Available from: <http://www.nci.edu.eg>.
- Emre, M. H., Aktay, G., Polat, A. and Vardt, N. 2007. Effects of benzo[a]pyrene and ethanol on oxidative stress of brain, lung tissues and lung morphology in rats. *Chin J Physiol*; 50:143-8.
- Gali-Muhtasib, H. U., Aboukheir, W., Kheir, L., Darwiche, N. and Crooks, P. 2004. Molecular pathway for Thymoquinone-induced cell cycle arrest and apoptosis in neoplastic keratinocytes. *Anticancer Drugs*, 15, 389-399.
- Gali-Muhtasib, H. Roessner, A. and Schneider-Stock, R. 2006. Thymoquinone: a promising anti-cancer drug from natural sources. *Int J Biochem Cell Biol*; 38:1249-53.
- Gardner, B.; Zhu, L.X.; Sharma, S.; Tashkin, D. P. and Dubinett, S.M. 2003. Methanandamide increases COX-2 expression and tumor growth in murine lung cancer, *FASEB J.*, 17(14), 2157-9.
- Gautam, S., Kwang, S., B. and Bharat, A. 2008. Targeting Nuclear Factor-kB Activation Pathway by Products and Enhancement of: Role in Suppression of Anti-apoptotic Gene. *Mol Cancer Res*, 6: 1059-1070.
- Gelboin, H. V. 1980. Benzo[a]pyrene metabolism, activation and carcinogenesis: role and regulation of mixed-function oxidases and related enzymes. *Physiol. Rev.* 60, 1107-1166.
- Hanspeter, W. Deuter, M. and Imilda, E. 2000. Chemoprevention of tobacco smoke induced lung carcinogenesis in mice. *Carcinogenesis*; 21: 977-82.
- Hecht, S.S., 1999. Tobacco smoke carcinogens and lung cancer. *J. Natl Cancer Inst.* 91, 1194-1210.
- Hecht SS, Upadhyaya P, Wang M, Bliss RL, McIntee E. J. and Kenney P. M. 2002. Inhibition of lung tumorigenesis in A/J mice by N acetyl S (N 2 phen-ethyl-thio-carbamoyl) Lcysteine and myoinositol, individually and in combination. *Carcinogenesis*; 23:1455-61.
- IARC 1989. Monographs on the Evaluation of the Carcinogenic Risk of Chemicals to Humans Diesel and Gasoline Exhausts and Some Nitroarenes, International Agency for Research on Cancer, Lyon, France.
- Kamara, S., Gopalakrishnan R., Pandi, A., Sundaram, J. and Thiruvengadam, D. 2009. Antioxidant and anticancer efficacy of hesperidin in benzo(a)pyrene induced lung carcinogenesis in mice *Investigational*



- New Drugs, Volume 27, pp. 214-222(9).
- Kamaraj, S., Anandakumar, P., Jagan, S., Ramakrishnan, G. and Devaki, T. 2007. Modulatory effect of hesperidin on benzo (a) pyrene induced experimental lung carcinogenesis with reference to COX2, MMP 2 and MMP9. *Journalhomepage:www.elsevier.com/locate/ejphar*
- Kanter, M., Coskun, O., Kalayci, M., Buyukbas, S. and Cagavi, F. 2006. Neuro-protective effects of *Nigella sativa* on experimental spinal cord injury in rats. *Hum Exp Toxicol*, 25: 127- 133.
- Kassie, F., Anderson, L. B. and Scherber, R. 2007. Indole-3-carbinol Inhibits 4-(Methyl-nitros-amino)-1-(3-pyridyl)-1-butanone Plus Benzo (a)pyrene Induced Lung Tumor-genesis in A/J Mice and Modulates Carcinogen-Induced Alterations in Protein Levels. *Cancer Res* 67:6502-6511.
- Kim, H.S., Kwack, S.J. and Lee, B.M. 2000. Lipid peroxidation, antioxidant enzymes, and benzo[a]pyrene-quinones in the blood of rats treated with benzo[a]pyrene. *Chem Biol Interact*; 127:139–50.
- Magesh, V., Durga, Bhavani K., Senthilnathan, P., Rajendran, P and Sakthisekaran, D. 2009. In Vivo Protective Effect of Crocetin on Benzo(a) pyrene-Induced Lung Cancer in Swiss Albino Mice. *Phytother. Res.* 23, 533–539.
- Mansour, M. A., Nagi, M. N., El-Khatib, A. S., and Al-Bekairi, A. M. 2002. Effects of thymoquinone on antioxidant enzyme activities, lipid peroxidation and DT-diaphorase in different tissues of mice: a possible mechanism of action. *Cell Biochemistry and Function*, 20, 143–151
- Mascaux, C., Martin, B., Verdebout, J.M., Ninane, V. and Sculier, J.P. 2005. COX-2 expression during early lung squamous cell carcinoma oncogenesis, *Eur. Respir. J.*: 26(2), 198-203.
- Mousavi, S.H., Tayarani-Najaran, Z., Asghari, M. and Sadeghnia, H.R. 2010. Protective Effect of *Nigella sativa* Extract and Thymoquinone on Serum/Glucose Deprivation-Induced PC12 Cells Death. *Cell Mol Neurobiol*, 30: 591- 598.
- Obrador, E., Carretero, J., Ortega, A., Medina, I., Rodilla, V. and Pellicer, J.A. 2002.  $\gamma$ -Glutamyl- Transpeptidase over-expression increases metastatic growth of B16 melanoma cells in the mouse liver. *Hepatology*; 35:74–81.
- Osann, K.E., Lowery, J.T. and Schell, M. J. 2000. Small cell lung cancer in women Risk associated with smoking, prior respiratory disease, and occupation. *Lung Cancer*; 28:1 10.
- Pagola, S., Benavente, A., Raschi, A., Romano, E., Molina, M. A., and Stephens, P. W. 2004. Crystal structure determination of thymoquinone by high-resolution X-ray powder diffraction. *AAPS Pharm Sci Tech*, 5, e28.
- Panandiker, A., Maru, G.B. and Rao, K.V.K. 1994. Dose response effects of malachite green on free radical formation, lipid peroxidation and DNA damage in hamster embryo cells and their modulation by antioxidants. *Carcinogenesis*; 15:2445–8.
- Ramakrishnan, G., Augustine, T.A., Jagan, S., Vinodhkumar, R. and Devaki, T. 2007. Effect of silymarin on N-nitroso-di-ethylamine induced hepatic-carcinogenesis in rats. *Exp Oncology*; 29:39–44.
- Rastogi, L., Feroz, S., Pandey, B.N., Jagtap, A. and Mishra, K.P. 2010. Protection against radiation-induced oxidative damage by an ethanolic extracts of *Nigella sativa* L. *NT J Radiat Biol*, 86: 719-731.
- Ren, W., Qiao, Z., Wang, H., Zhu L. and Zhang, L. 2003. Flavonoids Prom is anticancer agents. *Med Res Rev*; 23:519-34.

- Priya, D. K. Gayathri, R. Gunassekaran, G. R. Murugan and Sakthisekaran. 2011. Inhibitory effect of sulforaphane against Benzo(a)Pyrene induced lung cancer by modulation of biochemical signatures in female swiss albino mice. *Asian journal of biochemistry*
- Ruano-Ravina, A., Figueiras, A., Montes-Martínez, A. and Barros-Dios, J. M. 2003. Dose–response relationship between tobacco and lung cancer: new findings. *Eur. J. Cancer Prev.* 12, 257.
- Selvendiran, K. and Sakthisekaran, D. 2004. Chemo-preventive effect of piperine on modulating lipid peroxidation and membrane bound enzymes in benzo (a) pyrene induced lung carcinogenesis. *Biomed Pharmacother*; 58:264–7.
- Selvendiran, K., Senthilnathan, P., Magesh, V. and Sakthisekaran, D. 2004. Modulatory effect of Piperine on mitochondrial antioxidant system in Benzo (a) pyrene-induced experimental lung carcinogenesis. *Phyto medicine*; 11:85–9.
- Shaymaa, A. Abdel Atie (2014): Biochemical evaluation of novel nano composite in prevention of lung carcinoma in mice" Master thesis in veterinary Medical Sciences, (Biochemistry and Clinical Biochemistry), Faculty of Veterinary Medicine, Benha University, Egypt
- Shoieb, A. M., Elgayyar, M., Dudrick, P. S., Bell, J. L., Tit h of, P.K. 2003. In vitro inhibition of growth and induction of apoptosis in cancer cell lines by thymoquinone. *International Journal of Oncology*, 22, 107–113. [28].
- Sikkim, H, and Mulee, B. 2000. Lipid peroxidation and antioxidant enzymes in the blood of rats treated with benzo[a] pyrene. *Chem Biol Interact*; 127:139–50.
- Straif, K., R. Baan, Y. Grosse, B. Secretan, F. El Ghissassi and V. Cogliano. 2005. Carcinogenicity of polycyclic aromatic hydrocarbons, *Lance oncology*. 6 (12) (2005) 931–932.
- Wang ,H., Zang, W., Yang, Y., Zhang, Q., Zhao M., Gao Z., Li G., Meng, Q., Liu Q. and Zheng, X. 2013 .Tanshinone IIA and Baicalin inhibiting the formation of benzo[a]pyrene and benzo[a]pyrene induced cytotoxicity: Correlation with scavenging free radical. 403–410
- Wang, Z.F. 2009. Overexpression of Cu/Zn-superoxide dismutase and/or catalase accelerates benzo(a)pyrene detoxification by up regulation of the aryl hydrocarbon receptor in mouse endothelial cells. *Free Radic. Biol. Med.* 47, 1221–1229.
- Wester, P.W., 2012. Carcinogenic activity of benzo[a]pyrene in a 2 year oral study in Wistar rats. *Food Chem. Toxicology*. 50, 927–935.
- Winterhalder, R.C., Hirsch, F.R., Kotantoulas, G.K., Franklin, W.A. and Bunn P.A., 2004. Chemoprevention of lung cancer— from biology to clinical reality. *Ann. oncology*. 15:185.
- Worthen, D. R., Ghosheh, O. A. and Crooks, P. A. 1998. The in vitro Antitumor activity of some crude and purified components of black seed *Nigella sativa* L. *Anticancer Research*, 18, 1527–1532.
- Zhu, Y.M., Azahri, N.S., Yu, D.C. and Woll, P.J., 2008. Effects of COX-2 inhibition on-expression of vascular endothelial growth factor and interleukin-8 in lung cancer cells. *BMC Cancer* 8, 218.



## التأثير الكيميائي الوقائي للثيموكينون على سرطان الرئة المحدث بالبنزو(أ)بيرين في ذكور الجرذان

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### الملخص العربي

يعتبر الثيموكينون من المركبات التي لها تأثيرات مضادة للاكسده والالتهابات ضد العديد من الأمراض مثل مرض السكر والربو والتهاب المخ وسرطان الرئة. في هذه الدراسة تم دراسة تأثير الثيموكينون كمضاد للاكسده والالتهابات على العديد من العوامل الكيميائية كما تم دراسة استخدامه كعلاج للالتهابات وسرطان الرئة. في هذه الدراسة تم استخدام 100 من ذكور الجرذان حيث تم تقسيمها إلى أربع مجموعات رئيسيه. المجموعة الأولى ( المجموعة الضابطة ) :حيث تركت بلا علاج ، المجموعة الثانية (المجموعة المعالجة باستخدام البنزو(أ)بيرين ) حيث تم إعطاءها المادة المسرطنه (البنزو(أ)بيرين) عند جرعة 100 مجم /كيلوجرام من وزن الجسم وذلك لإحداث سرطان الرئة في ذكور الجرذان ، أما المجموعة الثالثة (مجموعة سرطان الرئة والمعالجة باستخدام الثيموكينون ) حيث تم إعطائها الثيموكينون عند جرعة 20 مجم / كيلوجرام من وزن الجرذان يوميا عن طريق الفم لمدة 8 أسابيع ، أما المجموعة الرابعة (مجموعة سرطان الرئة التي تم وقايتها باستخدام الثيموكينون ) حيث تم إعطائها الثيموكينون عند جرعة 20 مجم /كجم من وزن الجرذان يوم بعد يوم عن طريق الفم لمدة 30 أسبوع خلال فترة التجربة . تم اخذ عينات من الدم وأنسجة جسم الجرذان ( الرئة ) وقد أسفرت النتائج عن وجود انخفاض معنوي في إنزيم السوبر أكسيد ديسميوتيز والكتاليز ووجود زيادة في إل – مالون داي الدهيد بالاضافه إلى زيادة الهيبيتاجلوبين وإنزيمات السيكلواوكسيجيناز-2 والجاما جليوتوميل ترانسفيراز و تجزئة الحمض النووي دى ان ايه ودلالات الأورام كرسينوجينك امبريونك انتجين وأخيرا الكسباس 3 وقد أظهرت نتائج هذه الدراسة ان الثيموكينون كان له القدرة على علاج سرطان الرئة المحدث باستخدام البنزو(أ)بيرين وذلك من خلال الزيادة المعنوية في مضادات الاكسده السوبر أكسيد ديسميوتيز والكتاليز بالاضافه إلى النقص المعنوي في إل – مالون داي الدهيد بالاضافه إلى زيادة الهيبيتاجلوبين وإنزيمات السيكلواوكسيجيناز-2 والجاما جليوتوميل ترانسفيراز تجزئة الحمض النووي دى ان ايه ودلالات الأورام كرسينوجينك امبريونك انتجين وأخيرا الكسباس3. يمكن القول أن الثيموكينون قد يكون فعالا في الحد من سرطان الرئة بواسطة نشاطها وتأثيرها كمضاد للالتهابات، وتجديد آليات مضادة للأكسده الذاتية وانخفاض كاسباس 3 وتجزئة الحمض النووي في أنسجة الرئة. وتشير هذه النتائج أن كفاءة الثيموكينون كعامل وقائي متميز في الحد من سرطان الرئة.

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