



## QUALITY OF BEEF AND EDIBLE OFFAL AT ABATTOIR LEVEL

Hemmat, M. Ibrahim<sup>1</sup>, Reham, A. Amin<sup>1</sup>, Omima, A. Saleh<sup>2</sup>, El Shafay, M.S.

1 Food Control Dept., Fac. Vet. Med., Benha University. 2 Animal Health Research Institute, Damanhour Branch.

### ABSTRACT

A total of 120 random samples of cattle shoulder meat, liver, kidneys and lungs (30 of each) were collected from two traditional abattoirs of Elbehira province. All collected samples were subjected to organoleptic, chemical and microbiological examinations to determine their quality. The results showed that the sensory characters (color, odor and consistency) and chemical parameters (pH with the mean of  $5.70 \pm 0.04$ ,  $6.45 \pm 0.02$ ,  $6.49 \pm 0.01$ ,  $6.48 \pm 0.02$ , TVN with the mean of  $12.68 \pm 0.02$ ,  $13.06 \pm 0.04$ ,  $12.76 \pm 0.03$ ,  $12.98 \pm 0.04$  and TBA with the mean of  $0.24 \pm 0.01$ ,  $0.16 \pm 0.01$ ,  $0.25 \pm 0.01$ ,  $0.24 \pm 0.01$ ) for shoulder meat, liver, kidneys and lungs respectively were normal and accepted. On the other hand the results of microbiological examination in examined samples of shoulder meat, liver, kidneys and lungs revealed that the mean of total APC were  $2.36 \times 10^5 \pm 48 \times 10^3$ ,  $20.1 \times 10^4 \pm 37 \times 10^3$ ,  $3.43 \times 10^5 \pm 1.97 \times 10^5$ ,  $18.9 \times 10^4 \pm 3.8 \times 10^4$ , respectively. While the mean of *Enterobacteriaceae* count were  $10.8 \times 10^4 \pm 2.6 \times 10^4$ ,  $84 \times 10^3 \pm 18 \times 10^3$ ,  $69 \times 10^3 \pm 17 \times 10^3$ ,  $84 \times 10^3 \pm 21 \times 10^3$ , respectively, coliform count with the mean of  $44 \times 10^3 \pm 12 \times 10^3$ ,  $34 \times 10^3 \pm 7 \times 10^3$ ,  $22 \times 10^3 \pm 5 \times 10^3$ ,  $32 \times 10^3 \pm 8 \times 10^3$  respectively, total *Staphylococci* count with the mean of  $28 \times 10^3 \pm 5 \times 10^3$ ,  $23 \times 10^3 \pm 4 \times 10^3$ ,  $23 \times 10^3 \pm 5 \times 10^3$ ,  $20 \times 10^3 \pm 4 \times 10^3$  respectively, total mould with the mean of  $1.24 \times 10^2 \pm 0.64 \times 10^2$ ,  $0.46 \times 10^2 \pm 0.9 \times 10^2$ ,  $0.49 \times 10^2 \pm 0.1 \times 10^2$ ,  $0.87 \times 10^2 \pm 0.22 \times 10^2$ , respectively and total yeast count with the mean of  $2.59 \times 10^2 \pm 1.41 \times 10^2$ ,  $0.85 \times 10^2 \pm 0.36 \times 10^2$ ,  $0.23 \times 10^2 \pm 0.07 \times 10^2$ ,  $1.62 \times 10^2 \pm 0.8 \times 10^2$ , respectively, were higher than the permissible limits and the examined samples failed to be accepted.

**KEY WORDS:** Shoulder meat, liver, kidneys, lungs, organoleptic examination, chemical examination, microbiological examination, abattoirs.

(BVMJ-25(2): 254-263, 2013)

### 1- INTRODUCTION

Fresh meat is highly perishable due to its biological composition. The slaughter of animals yields many edible products other than carcass meat (such as red offal), which are fit for human consumption. They are used either as prepared items (e.g. slices of liver) or used as ingredients in meat products. The market for 'edible by-products' differs with country (even region) and culture (Devatkl *et al.*, 2004). The intact tissues of healthy slaughtered animals are mostly sterile but the meat may be contaminated during slaughtering, handling, processing and storage from hands, workers, clothes, knives, hide, gut, fecal material on feet or

from the environment. Microbial contamination of the carcass results in spoilage of meat, reduced shelf-life of meat and public health hazards (Phillips *et al.*, 2006) either due to presence of spoilage bacteria responsible for unfavorable changes or pathogenic bacteria leading to harmful effects as food infection or intoxication among consumers (Eley, 1992). Organoleptic, chemical and microbiological quality of fresh meat and edible offal have been receiving attention, all over the world, from researchers, food industry, health organization and governments due to the occurrence of significant outbreaks of food borne illness affecting consumers. Quality maintenance is important not only for consumer health

protection but also to assure uniformity in fresh meat shelf-life (Baumann-Popczyk and Sadkowska-Todays, 2012). So, the object of the current study was to evaluate the organoleptic, chemical and microbiological quality of cattle meat and edible offal at abattoir level.

### 2. MATERIAL AND METHODS:

#### 2.1. Collection of samples:

A grand total of 120 cut samples of cattle shoulder meat and internal edible offal (liver, kidneys and lungs) (30 of each) were equally collected from 30 different cattle carcasses slaughtered in two different traditional abattoirs in El Behera governorate (15 carcasses from each abattoir). The samples were collected after complete stamping of slaughtered animals, and transferred to the laboratory in an insulated ice box under complete aseptic conditions, without undue delay for organoleptic, chemical and microbiological examinations.

2.2. *Organoleptic Examination*: color, odor and consistency (Morr-Mary, 1970).

#### 2.3. Chemical Examination:

Determination of pH, TVN (FAO, 1980) and TBA (Kirk and Sawyers, 1991).

#### 2.4. Microbiological Examination:

- Determination of APC, *Enterobacteriaceae*, coliform and total *Staphylococci* counts (ICMSF, 1982).
- Determination of total mould and yeast count (Cruickshank *et al.*, 1975).
- Isolation and identification of mould and yeast (Refai, 1987).
- Isolation and identification of *Staphylococcus aureus* (ICMSF, 1996).

#### 2.5. Statistical analysis:

Data were analyzed by one way ANOVA. Means with different alphabetical superscripts in the same columns are significantly different at  $P \leq 0.05$ .

### 3. RESULTS:

From the results reported in table (1), it is obvious that 40%, 36.6% and 23.4% of the examined meat samples, 53.4%, 30% and 16.6% of the examined liver samples, 70%, 30% and zero% of the examined kidney samples and 43.4%, 26.6% and 30% of the examined lung samples took excellent, very good and good grades, respectively according to the quality system [1]. Regarding the results recorded in table (2), pH mean values  $5.70 \pm 0.04$  in the examined meat samples,  $6.45 \pm 0.02$  in the examined liver samples,  $6.49 \pm 0.01$  in the examined kidney samples and finally  $6.48 \pm 0.02$  in the examined lung samples. It is evident from the results recorded in table (2) that TVN mean values (mg/100gm)  $12.68 \pm 0.02$  in the examined meat samples,  $13.06 \pm 0.04$  in the examined liver samples,  $12.76 \pm 0.03$  in the examined kidney samples and finally  $12.98 \pm 0.04$  in the examined lung samples. Results achieved in table (2) revealed that TBA mean values (mg malonaldehyde/ kg of sample)  $0.24 \pm 0.01$  in the examined meat samples,  $0.16 \pm 0.01$  in the examined liver samples,  $0.25 \pm 0.01$  in the examined kidney samples and finally  $0.24 \pm 0.01$  in the examined lung samples. Moreover, table (2) revealed that there were high significant differences in pH, TVN and TBA values ( $p < 0.05$ ) between the examined samples of meat and edible offal. It is evident from the results recorded in table (3) that APC mean values (cfu/gm) in the examined samples  $2.23 \times 10^5 \pm 48 \times 10^3$  for shoulder meat,  $20.1 \times 10^4 \pm 37 \times 10^3$  for liver,  $3.43 \times 10^5 \pm 1.97 \times 10^5$  for kidneys and  $18.9 \times 10^4 \pm 3.8 \times 10^4$  for lungs. Table (3) indicated that the mean values of *Enterobacteriaceae* count (cfu/gm) in the examined samples  $10.8 \times 10^4 \pm 2.6 \times 10^4$ ,  $84 \times 10^3 \pm 18 \times 10^3$ ,  $69 \times 10^3 \pm 17 \times 10^3$  and  $84 \times 10^3 \pm 21 \times 10^3$  for shoulder meat, liver, kidneys and lungs respectively. From the obtained results recorded in table (3), it was clear that the mean values of coliform count (cfu/gm) in

the examined samples  $44 \times 10^3 \pm 12 \times 10^3$  for shoulder meat,  $34 \times 10^3 \pm 7 \times 10^3$  for liver,  $22 \times 10^3 \pm 5 \times 10^3$  for kidneys and  $32 \times 10^3 \pm 8 \times 10^3$  for lungs. The data recorded in table (3) revealed that the mean values of total *Staphylococci* count (cfu/gm) in the examined samples were  $28 \times 10^3 \pm 5 \times 10^3$  for shoulder meat,  $23 \times 10^3 \pm 4 \times 10^3$  for liver,  $23 \times 10^3 \pm 5 \times 10^3$  for kidneys and  $20 \times 10^3 \pm 4 \times 10^3$  for lungs. In other words, there were no significant differences in APC, *Enterobacteriaceae*, coliform and total *Staphylococci* counts ( $P < 0.05$ ) between the examined samples of meat and edible offal. Table (4) declared that 40%, 20%, 13.3% and 30% of the examined meat, liver, kidney and lung samples, respectively, were contaminated with *S. aureus*. It is evident from table (3) that the mean values of total mould count (cfu/gm) of the examined samples  $1.24 \times 10^2 \pm 0.64 \times 10^2$  for shoulder meat,  $0.46 \times 10^2 \pm 0.09 \times 10^2$  for liver,  $0.49 \times 10^2 \pm 0.1 \times 10^2$  for kidneys and  $0.87 \times 10^2 \pm 0.22 \times 10^2$  for lungs. Means within examined samples of meat and edible offal showed no significant differences ( $P < 0.05$ ). Identification of mould species isolated from the examined samples of meat and edible offal was shown in table (5). In shoulder meat were *Aspergillus spp.* 66.6%, *Penicillium spp.* 23.3%, *Geotrichum spp.* 43.3%, *Cladosporium spp.* 16.6%, *Fusarium spp.* 6.6%, *Alternaria spp.* 20% and *Mucor spp.* 36.6% but *Rhizopus spp.* failed to be detected, in liver were *Aspergillus spp.* 60%, *Penicillium spp.* 23.3%, *Geotrichum spp.* 13.3%, *Cladosporium spp.* 16.6%, *Fusarium spp.* 10%, *Alternaria spp.* 6.6%, *Rhizopus spp.* 10% and *Mucor spp.* 11%, in kidneys were *Aspergillus spp.* 56.6%, *Penicillium spp.* 13.3%, *Geotrichum spp.* 16.6%, *Fusarium spp.* 10%, *Alternaria spp.* 13.3%, *Rhizopus spp.* 6.6% and *Mucor spp.* 36.6%, but *Cladosporium spp.* failed to be detected and in lungs were *Aspergillus spp.* 63.3%, *Penicillium spp.* 20%, *Geotrichum spp.* 10%, *Cladosporium spp.* 23.3%, *Fusarium spp.* 20%, *Alternaria spp.* 16.6% and *Mucor spp.* 13.3% but *Rhizopus spp.*

failed to be detected. It is evident from table (3) that the mean values of total yeast count (cfu/gm) of examined samples  $2.59 \times 10^2 \pm 1.41 \times 10^2$  for shoulder meat,  $0.85 \times 10^2 \pm 0.36 \times 10^2$  for liver,  $0.23 \times 10^2 \pm 0.07 \times 10^2$  for kidneys and  $1.62 \times 10^2 \pm 0.8 \times 10^2$  for lungs. Means within examined samples of meat and edible offal showed no significant differences ( $P < 0.05$ ). Table (6) showed the incidence of species of yeast isolated from the examined samples of meat and edible offal. *Rhodotorulla* was detected in 50%, 56.6%, 33.3% and 53.3% of the examined meat, liver, kidney and lung samples, respectively. While, *Candida kiusci* was detected in 36.6%, 13.3%, 26.6% and 23.3% of the examined meat, liver, kidney and lung samples, respectively.

#### 4. DISCUSSION:

Meat and edible offal have long been considered as highly desirable, nutritious and protein-rich food, but at the same time, unfortunately, they are also highly perishable because they provide the nutrients needed to support the growth of many types of microorganisms. Due to their unique biological and chemical nature, their quality attributes deteriorate from the time of slaughter until consumption (Kalalou *et al.*, 2004). Due to lipid oxidation and bacterial growth which are the main factors that determine food quality loss and shelf life reduction. Lipid oxidation leads to the degradation of lipids and proteins which, in turn, contribute to the reduction in nutritional quality as well as deterioration in flavor, color and texture of displayed meat (Aguirrezábal *et al.*, 2000). Bacterial contamination can precipitate major public health hazards and economic losses in terms of food poisoning and meat spoilage (Fernández – López *et al.*, 2005). From the results reported in table (1), it is obvious that according to the quality system recommended by Devatkl *et al.* (2004). Accordingly, all the examined samples were accepted organoleptically. It could be concluded that the examined kidney

## Quality of beef and edible offal at abattoir level

Table (1): Organoleptic evaluation of examined cattle meat and offal samples at abattoir level (n= 30)

Samples	Meat			Liver		Kidneys		Lungs	
	Point	No.	%	No.	%	No.	%	No.	%
Excellent	10	12	40	16	53.4	21	70	13	43.4
Very good	9	11	36.6	9	30	9	30	8	26.6
Good	8	7	23.4	5	16.6	---	---	9	30

Table (2): Statistical analyses of chemical results of examined samples of cattle meat and edible offal at abattoir level (n=30)

Parameters	Meat	Liver	Kidneys	Lungs
PH	5.70 ± 0.04b	6.45 ± 0.02a	6.49 ± 0.01a	6.48 ± 0.02a
TVN	12.68 ± 0.02b	13.06 ± 0.04a	12.76 ± 0.03b	12.98 ± 0.04a
TBA	0.24 ± 0.01a	0.16 ± 0.01b	0.25 ± 0.01a	0.24 ± 0.01a

There were high significant differences ( $P < 0.05$ ) in pH, TVN and TBA values of the examined samples.

Table (3): Statistical analyses of microbiological results of examined samples of cattle meat and edible offal at abattoir level (n=30)

Count CFU/g	Meat	Liver	Kidney	Lung
APC	2.36×10 <sup>5</sup> ±48×10 <sup>3</sup> a	20.1×10 <sup>4</sup> ±37×10 <sup>3</sup> a	3.43×10 <sup>5</sup> ±1.97×10 <sup>5</sup> a	18.9×10 <sup>4</sup> ±3.8×10 <sup>4</sup> a
EC	10.8×10 <sup>4</sup> ±2.6×10 <sup>4</sup> a	84×10 <sup>3</sup> ±18×10 <sup>3</sup> a	69×10 <sup>3</sup> ±17×10 <sup>3</sup> a	84×10 <sup>3</sup> ±21×10 <sup>3</sup> a
CC	44×10 <sup>3</sup> ±12×10 <sup>3</sup> a	34×10 <sup>3</sup> ±7x10 <sup>3</sup> a	22x10 <sup>3</sup> ±5x10 <sup>3</sup> a	32x10 <sup>3</sup> ±8x10 <sup>3</sup> a
TSC	28×10 <sup>3</sup> ±5×10 <sup>3</sup> a	23x10 <sup>3</sup> ±4x10 <sup>3</sup> a	23x10 <sup>3</sup> ±5x10 <sup>3</sup> a	20x10 <sup>3</sup> ±4x10 <sup>3</sup> a
TMC	1.24×10 <sup>2</sup> ±0.64×10 <sup>2</sup> a	0.46×10 <sup>2</sup> ±0.09×10 <sup>2</sup>	0.49×10 <sup>2</sup> ±0.1×10 <sup>2</sup> a	0.87×10 <sup>2</sup> ±0.22×10 <sup>2</sup> a
TYC	2.59×10 <sup>2</sup> ±1.41×10 <sup>2</sup> a	a 0.85×10 <sup>2</sup> ±0.36×10 <sup>2</sup> a	0.23×10 <sup>2</sup> ±0.07×10 <sup>2</sup> a	1.62×10 <sup>2</sup> ±0.8×10 <sup>2</sup> a

There were no significant differences ( $P < 0.05$ ) in APC, TEC, TCC, TSC, TMC and TYC of the examined samples.

-APC: Aerobic Plate Count.

-EC: *Enterobacteriaceae* Count.

-CC: Coliform Count.

- TSC: Total Staphylococcal Count

-TMC: Total Mould Count.

- TYC: Total Yeast Count

Table (4): Incidence of *Staphylococcus aureus* isolated from the examined samples of cattle meat and edible offal at abattoir level (n=30)

Samples	No.	%
Meat	12	40
Liver	6	20
Kidney	4	13.3
Lung	9	30

Table (5): Incidence of mould species isolated from the examined samples of cattle meat and edible offal at abattoir level (n = 30)

Samples	Mould spp.	Meat		Liver		Kidneys		Lungs	
		No.	%	No.	%	No.	%	No.	%
	<i>Aspergillus spp.</i>	20	66.6	18	60	17	56.6	19	63.3
	<i>Penicillium spp.</i>	7	23.3	7	23.3	4	13.3	6	20
	<i>Geotrichum spp.</i>	13	43.3	4	13.3	5	16.6	3	10
	<i>Cladosporium spp.</i>	5	16.6	5	16.6	---	---	7	23.3
	<i>Fusarium spp.</i>	2	6.6	3	10	3	10	6	20
	<i>Alternaria spp.</i>	6	20	2	6.6	4	13.3	5	16.6
	<i>Rhizopus spp.</i>	---	---	3	10	2	6.6	---	---
	<i>Mucour spp.</i>	11	36.6	5	16.6	11	36.6	4	13.3

Table (6): Incidence of yeast species isolated from the examined samples of cattle meat and edible offal at abattoir level (n = 30)

Samples	Meat		Liver		Kidneys		Lungs	
	No.	%	No.	%	No.	%	No.	%
Mould spp.								
<i>Rhodotorulla</i>	15	50	17	56.6	10	33.3	16	53.3
<i>Candida kiusci</i>	11	36.6	4	13.3	8	26.6	7	23.3

samples showed superior organoleptic quality than the examined lung samples. Such findings may be attributed to the fact that kidneys are embedded in body fat and remain hanged in the body cavity not handled except by the veterinarian's knife. However, lungs undergo numerous faulty manipulations and handling from butchers. Regarding the results recorded in table (2), pH mean values in the examined samples and according to the safe permissible limit stipulated by EOS (2005) for pH in red meat (5.6 - 6.2) and edible offal (6 - 6.8), it was indicated that all the examined samples of meat and edible offal were in accordance with this limit. The obtained results were nearly similar to those reported by Immonen *et al.* (2000). While, higher results were obtained by El-Shamy (2011) in the examined liver samples ( $6.96 \pm 0.09$ ).

However, lower results were reported El-Shamy (2011) in the examined lung samples ( $6.08 \pm 0.07$ ). pH value plays an important role for the microbiological growth quality affecting the shelf life of meat (Hathout-Amal and Aly-Soher, 2010). It is evident from the results recorded in table (2) that TVN mean values showed that all the examined samples of meat and edible offal were accepted according to the safe permissible limit recommended by EOS (2005) for TVN in red meat (should not exceed 20 mg/100 gm) and edible offal (should not exceed 30 mg/100 gm). TVN value was more useful for assessing the degree of meat deterioration than for evaluating the changes occurring during the first storage stages (El Marrakchi *et al.*, 1990).

## Quality of beef and edible offal at abattoir level

Results achieved in table (2) revealed that TBA mean values (mg malonaldehyde/kg of sample) in the examined meat and edible offal were accepted based on their TBA content according to EOS (2005) which stated that the maximum permissible limit for TBA in meat and edible offal should not exceed 0.9 mg malonaldehyde/kg of sample. TBA is a good indicator of the quality of meat. TBA value is a widely used indicator for the assessment of degree of lipid oxidation (Raharjo and Sofos, 1993). It is evident from the results recorded in table (3) that the mean values of APC (cfu/gm) in the examined samples of meat and edible offal and according to the safe permissible limit stipulated by EOS (2005) for APC in red meat (not exceed  $10^6$  cfu/gm) and edible offal (not exceed  $10^5$  cfu/gm), it was indicated that all the examined samples of red meat were in accordance with this limit. While, all the examined samples of edible offal were not in accordance with this limit. Concerning red meat cuts, nearly similar results were obtained by Feizullah and Daskalov (2010). However, lower results were obtained by Shimaa (2012). While, higher results were obtained by Hejazi (2013). Regarding to edible offal, lower results were obtained by Ammar (2012), but higher results were obtained by Rasha (2013). Aerobic plate count is generally accepted as a criterion for microbial contamination of carcasses and a useful indicator of hygienic conditions of abattoir (Cohen *et al.*, 2007).

Table (3) indicated that the mean values of total *Enterobacteriaceae* count (cfu/gm) in the examined samples of meat and edible offal were unaccepted based on their *Enterobacteriaceae* count according to EC (2007) which stated that the maximum permissible limit for *Enterobacteriaceae* count in meat and edible offal should not exceed  $3.17 \times 10^2$  cfu/gm. Regarding to red meat, nearly similar results were obtained by Hejazi (2013). However, higher results were obtained by Ali (1992) and lower results were obtained by Feizullah and Daskalov (2010), Sabik (2011), and Shimaa

(2012). Concerning edible offal, higher results were obtained by El-Shamy (2011). While, lower results were obtained by Ammar (2012).

*Enterobacteriaceae* have an epidemiological importance as some of their members are pathogenic and may cause serious infections and food poisoning outbreaks to human being. The presence of *Enterobacteriaceae* in large numbers in food indicates improper hygienic measures, inadequate processing or recontamination due to cross contamination by raw materials, dirty equipment or unhygienic handling (Gill and Landers, 2004).

From the obtained results recorded in table (3), it was clear that the mean values of coliform count (cfu/gm) in the examined meat and edible offal and according to the safe permissible limit stipulated by FAM [33] for total coliform count in red meat (not exceed  $10^3$  cfu/gm) and edible offal (not exceed  $10^2$  cfu/gm), it was indicated that all the examined samples of red meat and edible offal were unaccepted with this limit. The current results of red meat were nearly similar with those obtained by Hejazi (2013). While, higher results were obtained by Yadav *et al.* (2006) and lower results were obtained by Sabik (2011) and Shimaa (2012)  $4.36 \times 10^2$  (cfu/gm). On the other hand, nearly similar results of edible offal were obtained by Ammar (2012). While, higher results were obtained by El-Shamy (2011). Furthermore, the high coliform count of edible offal may be attributed to the unsanitary conditions of offal collection after evisceration; putting offal on floor contaminated with fecal matters and delayed transportation of offal to special hygienic place. Total coliform count is used as general indicator of water pollution or sanitary conditions in the food processing environment (Feng *et al.*, 2002).

The data recorded in table (3) revealed that the mean values of total *Staphylococci* count (cfu/gm) in the examined samples of meat and edible offal nearly similar with results in red meat which obtained by Sabik (2011). However, lower results were

obtained by El-Shamy (2008). Higher results were obtained by Hejazi (2013). While, nearly similar results in edible offal were obtained by El-Shamy (2011). Meanwhile, lower results were obtained by Ammar (2012). Higher results were obtained by Rasha (2013). *Staphylococci* are commonly found in the skin and upper respiratory tract of man and animals and can easily contaminate the carcass. The presence of *Staphylococci* on carcass surface may be due to contamination during dressing and evisceration in slaughter house, contaminated equipment, butcher's hand with abrasions and wounds, slaughter of animal beside dressed one in the same area in the slaughter hall and contamination of air from crowdness of workers and their aerosols (Lasts *et al.*, 1992). The obtained results of red meat were nearly similar with those reported El-Shamy (2011). While, lower results were obtained by Sabik (2011) who mentioned the ratio was 4%. Concerning edible offal, lower results were obtained by Rasha (2013) who found coagulase positive *S. aureus* in 4% and 4% of the examined samples of beef liver and kidney, respectively.. Higher results were obtained by Ammar (2012) who found coagulase positive *S. aureus* in 42% and 28 % of the examined samples of beef liver and kidney, respectively.

*Staphylococcus aureus* enterotoxins are the predominant cause of gastrointestinal symptoms observed during intoxications. *Staphylococcus aureus* is considered the third most important cause of disease in the world amongst the reported food-borne illnesses (Tamarapu *et al.*, 2001).

It is evident from table (3) that the mean values of total mould count (cfu/gm) of the examined samples of meat and edible offal showed that all the examined samples of meat and edible offal were rejected based on their total mould count according to EOS (2005) which stated that meat and edible offal should be free from any fungal growth. Nearly similar results in red meat were obtained by Hejazi (2013). While, nearly similar results in edible offal were obtained

by El-Shamy (2011). Higher results were obtained by Rasha (2013) who mentioned that the average mould counts were  $2.97 \times 10^5$ ,  $1.04 \times 10^6$  and  $1.55 \times 10^5$  (cfu/gm) in the examined liver, kidney and lung samples, respectively.

Presence of mould in the examined samples may be attributed to the fact that mould need moisture to grow. So, they often found in environment as abattoir in which water is the base of the work (EL-Shamy, 2011). Mould count is used as an index of proper sanitation and high quality products. Mould can grow over an extremely wide range of temperature. They can assist in the putrefactive processes and may produce toxic substances namely mycotoxins which may lead to hemorrhages with hepatotoxic, carcinogenic or immunosuppressive effects (Hassan *et al.*, 2004). Identification of mould species isolated from the examined samples of meat and edible offal was shown in table [5]. These results when compared with another results obtained by El-Shamy (2011) and Rasha (2013) we found numerous variations in the rate of incidence and distribution of mould species on the examined samples. Some were agreed, some were lower and some were higher.

It is evident from table (3) that the mean values of total yeast count (cfu/gm) of examined samples of meat and edible offal showed no significant differences ( $P < 0.05$ ). Nearly similar results were obtained by El-Shamy (2011). Yeasts normally play a small role in spoilage because they constitute only a small portion of the initial population. They grow slowly in comparison with most bacteria and their growth may be limited by metabolic substances produced by bacteria. Spoilage yeasts find their way into food resulting in undesirable changes in physical appearance of food. Some species of yeast constitute a public health hazard as some species of *Candida* may cause gastrointestinal disturbances, vulvovaginitis, endocarditis, pulmonary infection, and occasionally fatal systemic disease (Jesenska and Hardinovva, 1981).

**5. REFERENCES**

1. Aguirrezábal, M.M., Mateo, J., Dominguez, M.C., Zumalacarregui, J.M. 2000. The effect of paprika, garlic and salt on rancidity in dry sausages. *J. Meat Sci.*, 54: 77-81.
2. Ali, M.H.I. 1992. Hygienic quality of mutton carcasses in Sharkia province. M. V. Sc. Thesis (Meat Hygiene), Fac.Vet.Med., Zagazig University.
3. Ammar, S. 2012. Microbiological quality of beef liver and kidney in Kafr El Sheikh. M. V. Sc., Thesis (Meat Hygiene), Fac. Vet. Med., Kafr El Sheikh University.
4. Baumann-Popczyk, A., Sadkowska-Todays, M. 2012. Food borne infection and intoxications in Poland in 2010. *Przegl Epidemiol.*, 66 (2): 241-248.
5. Cohen, N., Ennaji, H., Bouchrif, B., Hassar, M., Karib, H. 2007. Comparative study of microbiological quality of raw poultry meat at various seasons and for different slaughtering processes in Casablanca (Morocco). *J. Appl. Poul. Res.*, 16: 502–508.
6. Cruickshank, R., Duguid, J.P., Marino, B.P. and Swain, R.H.A. 1975. *Medical Microbiology*. 12th Ed. Vol., Churchill Livingstone, London and New York.
7. Devatkl, S., Mendiratta, S.K., Kondaiah, N., Sharma, M.C., Anjaneyulu, A.S.R. 2004. Physicochemical, functional and microbiological quality of buffalo liver. *J. Meat Sci.*, 68: 79-86.
8. Egyptian Organization for Standardization and Quality Control. “EOS”. 2005. Egyptian Organization for Standardization and Quality Control. No. 1522 for frozen meat.
9. Egyptian Organization for Standardization and Quality Control. “EOS”. 2005. Egyptian Organization for Standardization and Quality Control. No. 1473 for frozen liver.
10. Egyptian Organization for Standardization and Quality Control. “EOS”. 2005. Egyptian Organization for Standardization and Quality Control. No. 2062 for frozen kidneys, heart, spleen, brain, pancreas and tongue.
11. El Marrakchi, A., Bennour, M., Bouchriti, N., Hamama A., Tagafait, H. 1990. Sensory, chemical and microbiological assessment of Moroccan sardines (*Sardina pilchardus*) stored in ice. *J. Food Protec.*, 53: 55.
12. Eley, A. R. 1992. *Microbial food poisoning*. 1st Ed. P.46-51. Champan and Hall Publisher, London.
13. EL-Shamy, R.H.M. 2008. Microbial evaluation of slaughtering steps of food animal at Alexandria abattoir. M.V.Sc., Thesis (Meat Hygiene), Fac. Vet. Med. Alexandria University.
14. EL-Shamy, R.H.M. 2011. Quality assurance of internal edible offal produced from food animals abattoirs in Alexandria. Ph. D. Vet. Sc. Thesis (Meat Hygiene), Fact. Vet. Med., Alexandria University.
15. European Commission “EC”. 2007. Commission regulation (EC) No 1441/2007 of 5 December 2007. ISO 21528-2.
16. Feizullah, F., Daskalov, H. 2010. Investigation on lamb meat production hygiene in facilities with low and high production capacity. *Bulg. J. Vet. Med.* 13 (4): 252–258.
17. Feng, P., Weagent, S.D., Grant, M.A. 2002. Bacteriological Analytical Manual. Online. [www.lib.ncsu.edu/pubweb/www/ETDdb/web\\_root/collection/available/etd-04102005-213953/unrestricted/etd.pdf](http://www.lib.ncsu.edu/pubweb/www/ETDdb/web_root/collection/available/etd-04102005-213953/unrestricted/etd.pdf).
18. Fernández – López, J., Zhi, N., Aleson-Carbonell, L., Pérez- Alvarez, J. A. Kuri, V. 2005. Antioxidant and antibacterial activities of natural extracts: Application in beef meat balls. *J. Meat Sci.*, 69: 371- 380.
19. Food Administration Manual “FAM”. 1995. Microbiological Reference



- Criteria for Food, S. 11: Microbiological Criteria Version 2.0 October 1995. Page 19.
20. Food and Agriculture Organization "FAO" 1980. Manual of Food Quality Control. FAO, United Nation, Rome, Italy.
  21. Gill, C.O., Landers, C. 2004. Proximate sources of bacteria on boneless loins prepared from routinely processed and detained carcasses at a pork packing plant. *Int. J. Food Microbiol.*, 97: 171-178.
  22. Hassan, A. A., Ragheb, R. R., Rahmy, Nariman, A. 2004. Pathological changes in cows spontaneously fed on some mycotoxins. *Egypt. J. Comp. Path. and Clinic. Path.*, 17 (1): 282- 293.
  23. Hathout-Amal, S., Aly-Soher, E. 2010. Role of lactic acid bacteria as a biopreservative agent of Talbina. *J. American Sci.* 6: 889-898.
  24. Hejazi, M. 2013. Microbial changes in cattle carcasses stored at chilling condition. M.V.Sc., Thesis (Meat Hygiene), Fac.Vet.Med., Alexandria University.
  25. Immonen, K., Ruusunen, M., Puolanne, E. 2000. Some effects of residual glycogen concentration on the physical and sensory quality of normal pH beef. *J. Meat Sci.*, 55: 33-38.
  26. International Commission and Microbiological Specification for Foods "ICMSF" 1982. Microorganisms in foods. 2nd Ed., University of Toronto Press, Toronto, 188-192.
  27. International Commission and Microbiological Specification for Foods "ICMSF" 1996. Salmonellae. In: Roberts, T.A., Baird-Parker, A.C. Tompkin, R.B., (Eds.) Microorganisms in foods 5: Microbiological specifications of food pathogens. 1st Ed, Blackie Academic and Professional, London, UK, pp: 217-264.
  28. Jesenska, Z., Hardinova, I. 1981. Socialistic republic's lebensm unters forsch. 173 (1): 16- 20.
  29. Kalalou, I., Faid, M., Ahami, A.T. 2004. Extending the shelf life of fresh minced camel meat at ambient temperature by *Lactobacillus delbruekii* subsp. *delbruekii*. *Electronic J. Biotechnol.*, 7: 246-251.
  30. Kirk, R.S., Sawyers, R. 1991. Pearson's Composition and analysis of foods. 9th Ed. Longman, Scientific and technical London, UK.
  31. Lasts, J. A., Rodriguz, R., Zanelli, M., Margaria, A. 1992. Bacterial count from bovine carcass as an indicator for hygiene at slaughtering places. A proposal for sampling. *J. Food Protec.*, 54: 271.
  32. Morr-Mary, 1970. Introduction Food laboratory manual of food preparation and evaluation 2nd Ed. Mac Millan publishing Co., Newyork. Goolrer. Mac Millan publishing London.
  33. Phillips, Jordan, D., Morris, S., Jenson, I., Sumner, J. 2006. A national survey of the microbiological quality of beef carcasses and frozen boneless beef in Australia. *J. Food Protec.*, 69 (5): 1113-1117.
  34. Raharjo, S. J. N., Sofos, 1993. Methodology for measuring malonaldehyde as a product of lipid peroxidation in muscle tissues. *J. Meat Sci.* 35: 145- 169.
  35. Rasha, W.M.G. 2013. Microbial evaluation of some edible offal in bovine carcasses. M. V. Sc., Thesis (Meat Hygiene), Fac. Vet. Med., Benha University.
  36. Refai, M. 1987. Isolation and identification of fungi. Fac. Vet. Med., Cairo University.
  37. Sabik, I. 2011. Some bacterial food borne pathogen in cattle, camel and sheep carcasses. M. V. Sc. Thesis (Meat Hygiene), Fac. Vet. Med., Benha University.
  38. Shimaa, M.E. 2012. Detection of *Enterobacteriaceae* in meat and poultry cuts by using recent techniques. M. V. Sc., Thesis (Meat Hygiene), Fac.Vet. Med. Benha University.

39. Tamarapu, S., McKillip, J.L. and Drake, M. 2001. Development of a multiplex Polymerase chain reaction assay for detection and differentiation of *S. aureus* in dairy products. *J. Food Protec.*, 64: 664–668.
40. Yadav, M.M., Tale, S., Sharda, R., Sharma, V., Tiwari, S., Garg, U.K. 2006. Bacteriological quality of sheep meat in Mhow town of India. *Int. J. Food Sci. Technol.*, 41: 1234–1238.

### جودة لحوم الأبقار وأحشائها على مستوى المجازر

همت مصطفى إبراهيم<sup>1</sup> ريهام عبد العزيز أمين<sup>1</sup> أميمة عبد الفتاح صالح<sup>2</sup> مجدي صابر الشافعي<sup>1</sup>  
 أقسم مراقبة الأغذية - جامعة بنها - كلية الطب البيطري،<sup>2</sup>معهد بحوث صحة الحيوان- فرع دمنهور

### الملخص العربي

اجريت هذه الدراسة بمجازر محافظة البحيرة على ذبائح الأبقار حيث تم اخذ 120 قطعة عينة من اللحم والاحشاء وذلك بمعدل 30 عينة من لحم الكتف و30 عينة من الكبد و30 عينة من الكلى و30 عينة من الرنتين لمعرفة التغيرات الحسية مثل اللون والرائحة والقوام وكانت كلها طبيعيه وذات جوده عاليه. كما تم عمل بعض الاختبارات الكيمياءيه مثل قيمة الايدروجين في عينات اللحوم هو 5.7 والكبد 6.45 والكلى 6.49 والرنتين 6.48. ثانيا نسبة المركبات النيتروجين الطياره وكان نسبة متوسط نتائجها (مجم/100جم) في عينات اللحوم والكبد والكلى والرنتين على الترتيب هو 12.68 ، 13.06 ، 12.76 ، 12.98. ثالثا نسبة حامض الثيوباربيتينورك (مجم مالونالدهيد/كجم عينه) وكان نسبة متوسط نتائجها في عينات اللحوم والكبد والكلى والرنتين على الترتيب هو 0.24 ، 0.16 ، 0.25 ، 0.24. تم عمل الاختبارات الميكروبيه للحوم والاحشاء مثل العد الكلى للبكتيريا الهوائية وكان متوسط العدد الكلى للبكتيريا الهوائية في عينات اللحوم والكبد والكلى والرنتين على الترتيب هو  $2.36 \times 10^5$  ،  $20.1 \times 10^4$  ،  $3.43 \times 10^5$  ،  $18.9 \times 10^4$  لكل جم والعد الكلى للبكتيريا المعويه وكان متوسط العدد الكلى للبكتيريا المعويه في عينات اللحوم والكبد والكلى والرنتين على الترتيب هو  $10.8 \times 10^4$  ،  $84 \times 10^3$  ،  $69 \times 10^3$  ،  $84 \times 10^3$  لكل جم والعد الكلى للبكتيريا القولونية وكان متوسط العد الكلى للبكتيريا القولونية في عينات اللحم والكبد والكلى والرنتين على الترتيب هو  $44 \times 10^3$  ،  $34 \times 10^3$  ،  $22 \times 10^3$  ،  $32 \times 10^3$  لكل جم والعد الكلى للبكتيريا العنقوديه وكان متوسط العدد الكلى للبكتيريا العنقوديه في عينات اللحم والكبد والكلى والرنتين على الترتيب هو  $28 \times 10^3$  ،  $23 \times 10^3$  ،  $23 \times 10^3$  ،  $20 \times 10^3$  لكل جم. وقد تم عزل بكتيريا العنقود الذهبى من عينات اللحم والكبد والكلى والرنتين بنسب 40% ، 20% ، 13% ، 30% على التوالي. العد الكلى للخميره الفطريه وكان متوسط العدد الكلى للخميره في عينات اللحم والكبد والكلى والرنتين على الترتيب هو  $2.59 \times 10^2$  ،  $0.85 \times 10^2$  ،  $0.23 \times 10^2$  ،  $1.62 \times 10^2$  لكل جم. وقد تم عزل خميرة رودوتوريولا من عينات اللحم والكبد والكلى والرنتين بنسب 50% ، 57% ، 33% ، 53% على التوالي وايضا تم عزل خميرة كانديدا كويسكى من عينات اللحم والكبد والكلى والرنتين بنسب 37% ، 13% ، 27% ، 23% على التوالي. العد الكلى لفطريات العفن وكان متوسط العدد الكلى للعفن في عينات اللحم والكبد والكلى والرنتين على الترتيب هو  $1.24 \times 10^2$  ،  $0.46 \times 10^2$  ،  $0.49 \times 10^2$  ،  $0.87 \times 10^2$  لكل جم. وقد تم عزل انواع من العفن فى عينات اللحم وجد انه نسبة الاسبيرجيلس ، البنسيليوم ، الجيوتريك ، الكلادوسبوريم ، الفيوزاريم ، الالثيرناريا ، الميوكر هي 66.6% ، 23.3% ، 43.3% ، 16.6% ، 6.6% ، 20% ، 36.6 على التوالي ولا وجود للريزوبس. وفى عينات الكبد وجد ان نسبة الاسبيرجيلس ، البنسيليوم ، الجيوتريك ، الكلادوسبوريم ، الفيوزاريم ، الالثيرناريا ، الريزوبس ، الميوكر هي 60% ، 23.3% ، 13.3% ، 16.6% ، 10% ، 10% ، 6.6% ، 10% ، 16.6% على التوالي. بينما كانت فى الكلى نسبة الاسبيرجيلس ، البنسيليوم ، الجيوتريك ، الفيوزاريم ، الالثيرناريا ، الريزوبس ، الميوكر هي 56.6% ، 13.3% ، 16.6% ، 10% ، 10% ، 13.3% ، 6.6% ، 36.6% على التوالي. اما فى عينات الرنتين كانت نسبة الاسبيرجيلس ، البنسيليوم ، الجيوتريك ، الكلادوسبوريم ، الفيوزاريم ، الالثيرناريا ، الميوكر هي 63.3% ، 20% ، 10% ، 23.3% ، 20% ، 16.6% ، 13.3% على التوالي ولا وجود للريزوبس. وقد خلصت هذه الدراسة الى خطورة تلوث اللحوم والاحشاء بالجراثيم المختلفه والتي لها تاثير ضار على الصحة العامه.

(مجلة بنها للعلوم الطبية البيطرية: عدد 25(2):254-263, ديسمبر 2013)