

EPIZOTIOLOGICAL STUDIES ON PROLIFERATIVE KIDNEY DISEASE IN TILAPIA (OREOCHROMIS NILOTICUS) AND AFRICAN CATFISH (CLARIAS GARIEPINUS)

Eman I. Soror^a, Karima F. Mahrous^b, Ismail A.M.^c, Amany A. Abbass^a

^aDept. Fish Diseases and Management, Fac. Vet. Med., Benha Univ., ^bDept. Cell Biology, National Research Center, ^cDept. Fish Diseases and Management, Fac. Vet. Med., Suez Canal Univ.

A B S T R A C T

The proliferative kidney disease (PKD) has been documented to cause particular economic loss in fish farms worldwide. The present investigation aims to determine the prevalence and the etiology of (PKD) in Oreochromis niloticus and Clarias gariepinus obtained from El-Riah El-Tawfiki and its tributaries. To achieve this goal, 500 fishes were used of which 266 were O. niloticus and 234 were C. gariepinus. The fishes were dissected and the kidneys were examined macroscopically for the presence of morphological abnormalities and nodules. Fresh and Giemsa-stained slides of kidney tissue were also examined for determination of the causative agent. The results showed the prevalence of PKD in O. niloticus was higher in autumn (95.08%) and spring (91.94%) than in summer (76 %) and winter (73.53%). The prevalence of PKD in C. gariepinus was high in winter season (76%) and low in spring season (36.84 %). The overall prevalence of PKD was higher in O. niloticus (83.46%) than in C. gariepinus (46.58%). The Clinical signs of fish affected with PKD may be non-specific included distended abdomen with dark color, anaemia with pale gills and emaciation. Some fish have nodules in the eve around the iris forming a ring. Macroscopically, there was enlargement of the kidney with appearance of some kidney nodules of approximately 0.2 - 0.6 mm diameter. Based on the spore morphology, the causative agent was identified as different types of Myxosprean spores in O. niloticus and C. gariepinus. The high prevalence of PKD in both types of fishes suggests the need to establish strict control measure to overcome the great economic losses imposed by the disease.

KEY WORDS: Clarias gariepinus, Myxosporean, Oreochromis niloticus, PKD, Prevalence.

(BVMJ-23 [1]: 150-158, 2012)

1. INTRODUCTION

roliferative kidney disease (PKD) is serious disease principally a affecting populations of farmed and wild salmonid fish [16]. The disease is characterized by a severe swelling of the kidney induced by the host immune response the to presence of extrasporogonic stages of a Myxozoan parasite [8, 18, 26]. PKD is an example of devastating disease which is a result of infection by a Myxozoan parasite. Although the organism causing PKD was recognized for many decades it was only determined to be a Myxozoan in the 1980s

[17]. The recent discovery based on molecular evidence that freshwater Bryozoans are hosts for the causative agents of PKD [4]. The parasite was described as *Tetracapsuloid bryosalmonae* and finally identified the long sought source of PKD which is not transmitted from fish to fish [7]. PKD has been documented to cause particular economic loss to rainbow trout fish farms in the UK and Europe and to salmon hatcheries in North America [16]. PKD is also a disease of wild and feral fishes [13, 22, 32] and appears to be on the increases possible as a sequence of increasing temperatures [11, 32]. In Egypt, proliferative kidney disease was observed to be epidemic among Tilapian fishes and known as Tilapian proliferative kidney disease (TPKD) [10]. The disease is characterized by a severe swelling of the kidney induced by the host immune response to the presence of extrasporogonic stages of a Myxozoan parasite [8, 18, 26]. The disease is manifested as a massive immune response presence of **Tetracapsuloid** to the but **Bryosalmonae** the temperature dependence of the fish's immune systems means that the disease does not develop until temperatures exceed 15 °C although fishes may be infected at lower temperatures [15]. Because of the serious and the apparently growing impact of PKD in farmed and wild fishes, this study was conducted to investigate the prevalence of PKD in Oreachromus niloticus and *Clarias gariepinus*. The clinical and macroscopical findings were recorded. The morphology of disease-causing spores was presented.

2. MATERIAL AND METHODS

2.1. Fish

In this study 500 fish were investigated, of which 266 Oreochromis niloticus with average weight of 120±10 g and 234 Clarias gariepinus with average weight of 200±15g. The fishes were collected from El-Riah El-Tawfiki and its tributaries during the period from March 2010 to August 2011. The fishes were transmitted alive in large plastic containers with sufficient quantity of water obtained from where the fishes were collected. The freshly dead fishes were labeled and packed in clean plastic bags and kept in an ice boxer. The collected fishes were transported as rapid as possible to the laboratory of department of fish diseases and management, faculty of veterinary medicine, Benha University where clinical parasitological examination were and

conducted. Freshly dead fishes were subjected to immediate examination.

2.2. Aquaria

Clean glass aquaria measured $100 \times 30 \times 50$ cm were used for holding the fishes during the examination. The Aquaria were supplied with sufficient aeration by using electric air pump. The fishes were fed on pelleted commercial ration containing 25 % crude protein.

2.3. *Clinical examination*:

The fishes were examined externally for any abnormalities as previously described by Noga [24].

2.4. Postmortem examination

Dissection of fish and post-mortem examination for any internal abnormalities of the body cavity, kidneys, and urinary bladders were done according to Noga [24]. Particular attention was given to examination of kidney for any change in size, colour or presence of nodules or cysts.

2.5. Microscopic examinations of kidney preparations

Specimens from examined kidneys and nodules if present with few drops of saline were squashed and examined under microscope. In addition, air-dried smears fixed with methanol and stained with Giemsa used for further examinations [21]. Images were captured using Sony digital camera (SDC-p92, Sony Corporation).

2.6. Statistical analysis

The prevalence of PKD in both *O. niloticus* and *C. gariepinus* in different seasons was determined as the percentage of the affected fish from the total population examined using excel program.

3. RESULTS

3.1. The Prevalence of PKD

The prevalence of PKD in *O. niloticus* was higher in autumn (95.08%) and spring (91.94%) and lower at winter season

(73.53%). Moderately low infection was described in summer season (76%). On the other hand, the prevalence of PKD infection in *C. gariepinus* was high in winter season (76%) and low in spring

(36.84 %), summer (38.71 %) and autumn (41.30 %) seasons. The overall prevalence of PKD in *O. niloticus* (83.46%) was higher than the *C. gariepinus* (46.58%) (Table1, Fig.1).

Table 1 Seasonal prevalence of PKD in Oreochromis niloticus and Clarias gariepinus

Season	Oreochromis niloticus			Clarias gariepinus		
	Examined	Diseased	Incidence (%)	Examined	Diseased	Incidence (%)
Spring	62	57	91.94	76	28	36.84
Summer	75	57	76.00	62	24	38.71
Autumn	61	58	95.08	46	19	41.30
Winter	68	50	73.53	50	38	76.00
Totals	266	222	83.46	234	109	46.58

3.2. Clinical signs of PKD

The clinical signs may be absent or unspecific. Some affected fishes showed distended abdomen with dark color (Fig. 2), exophthalmos, appeared anaemic with pale gills and emaciation. Some fishes have nodules in the eye around the iris forming a ring (Fig. 3) and some fish appeared to be normal with no pathognomonic external lesions or signs. However. microscopical and histopathological examination of the kidneys of these apparently normal fish revealed various Myxosporeans.

3.3. Macroscopical examination

All the affected fishes showed enlargement of the kidneys (Fig. 4) and spleen (Fig. 5) and ascites but the pathognomonic PM lesion was the enlargement of the posterior kidney that may be protruded ventrally toward the abdominal cavity (Fig. 6) and sometimes distended urinary bladder. Also, we found macroscopic yellowish white nodules (2-3 mm or more in diameter) mostly appeared at the underlying side of the kidney in O. niloticus where at the surface of kidney in C. gariepinus (Fig. 7 8). When these nodules were and examined it found to contain myxosporean spores.

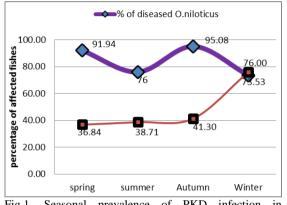


Fig.1. Seasonal prevalence of PKD infection in oreochromus niloticus and Clarias gariepinus.



Fig.2. Distended abdomen with dark color in *Oreochromis niloticus* affected with PKD



Fig. 3 A tilapia fish with PKD have nodules in the eye around the iris forming a ring.



Fig. 4. Enlargement of the kidney in *Clarias gariepinus* affected with PKD



Fig. 5. Enlargement of kidneys and spleen in Oreochromis *niloticus* affected with PKD.



Fig. 6. Enlargement of the kidney that is protruded ventrally toward the abdominal cavity in *Oreochromis niloticus* affected with PKD



Fig. 7. Enlargement of the kidney with macroscopic yellowish white nodules of different sizes in *Clarias gariepinus* affected with PKD.

3.4. Microscopical examination

Different types of myxosporean spores were identified by microscopical examination of the fresh preparations of affected kidneys. The infection demonstrated as mixed types of spores and almost no single infection within the fish observed (Fig. 9). Microscopic nodules were also observed in some affected cases. These nodules were either surrounded by intact capsule or the capsule was ruptured with release of the spores (Fig. 10)



Fig. 8. Macroscopic white nodules of different sizes in the kidney of *Clarias gariepinus* affected with PKD.

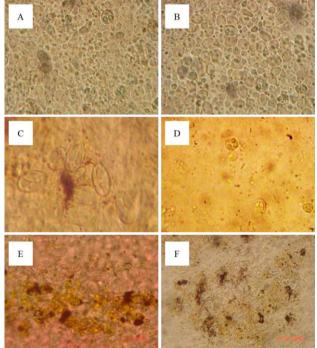


Fig.9. Fresh preparations of kidneys affected with PKD examined by microscopic examination showing different Myxosporean spores (A-F). All preparation showed mixed types of spores within the same slide.

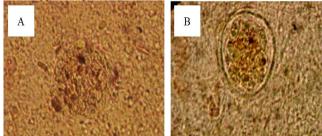


Fig. 10. Fresh preparations of kidneys affected with PKD showing microscopic nodules.

Different types of spores of myxosporea were identified in Giemsa-stained preparation as shown in Fig. 11. The spores have different sizes and morphology. However, in all stained preparation, the polar bodies are densely stained than the sporoplasm.

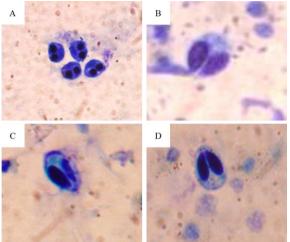


Fig. 11. Stained kidney tissue preparation (A-D) showing different myxosporean spores of varying sizes and morphology. Notice that the polar bodies are densely stained than the sporoplasm.

4. DISCUSSION

This study demonstrated that the overall prevalence of PKD in O. niloticus was 83.64%. This result coincided with that reported by Eissa et al. [9] who reported that heavy infection with Myxosoma spp. was recorded among cultured O. niloticus from Sharkiya governorate with prevalence infection exceeded 80 % of all examined fish. The high prevalence (more than 80 %) of Myxosporean infection among the examined fish highly suggests that such infection is endemic in the ponds used for rearing of these fish. This result is also comparable to that observed by Feist and Longshaw [12] who recorded that PKD is highly problematic for fish farms and hatcheries where up to 100% of stock can be infected. In consistent with this result, Okamura et al. [25] observed that PKD prevalence was variable range from 0 to 90-100%. On the other hand, Shehab El-Din [28] demonstrated a lower rate of infection (10.56%) in tilapian fish. The

prevalence of PKD in O. niloticus was higher in autumn ((95.08%) and spring (91.94%) and lower at winter season (73.53%). These results coincided with those reported by Hedrick *et al.* [16]. They reported that the PKD is often seasonally dependent occurring at water temperature above 15 °C in the fall months of the year. These results were also confirmed by higher prevalence during the month of October and November which are considered as the fall months in Egypt. Gay et al. [15] demonstrated that the disease does not develop until temperature exceeds 15 °C. Therefore, the disease prevalence will be reduced during the winter season, a result that coincided with the result of this study where the lowest prevalence of the disease was observed in winter season. The occurrence of PKD is associated with seasonal increase in water temperature above 15 °C [8]. In addition, the seasonal elevation of water temperature causes reduction of water dissolved oxygen and increases the susceptibility to the disease. The results of this study disagree with that reported by Tops et al. [30] and Tops and Okamura [31] who mentioned that the peak prevalence of PKD in rainbow trout Oncorhynchus mykiss in the fish farm occurred in summer (up to 100%). This can be explained by the difference in temperature between Europe and Egypt where the summer temperature in Europe is significantly lower than that in Egypt. In this study, a lower prevalence of PKD was observed in the summer season (76 %) compared to autumn (95.08%) and spring (91.64%). A more recent study in rainbow trout Oncorhynchus mykiss demonstrated that at 18°C, fish affected with PKD showed a gradual recovery of normal kidney morphology which was associated with a decline in parasite numbers and infection prevalence than fish kept at 12°C [27]. This implies that the increase of water temperature at 18°C or above has a negative effect on the disease prevalence. The common consistent finding with previous studies was that the prevalence increase in warmer temperature regions [29] as the temperature promotes disease development, enhances bryozoan biomass and increases spore production [25]. The discrepancies in the prevalence rate between this study and other studies United conducted in Kingdom demonstrated by Tops et al. [30] suggested that other factors rather than water temperature could have an effect on the prevalence of the PKD disease. This is particularly true since some previous demonstrated studies on PKD the existence of strong correlation between organic pollution of water, the presence of bryozoa and the outbreak of PKD [11]. In addition. Wahli et al. [32] reported that water contaminants can influence the PKD via development of the parasite in the bryozoan host or in its fish host. Therefore, the factors affecting prevalence of PKD in O. niloticus in Egypt may require further investigations and studies.

The prevalence of PKD infection in C. gariepinus was high in winter season (76%) and low in spring (36.84%), summer (38.71 %) and autumn (41.30 %) seasons. The overall prevalence of PKD in C. gariepinus was 46.6%. These findings partially agreed with those reported by Abbass et al. [1] who found that the prevalence of Myxozoan infestation in C. gariepinus was 42.5% and the maximum prevalence was observed in the winter (70%) and spring (80%) seasons. The results also agreed with Abdel-latif [2] who demonstrated that the highest prevalence of parasitic gill infestation by Myxosporeans in C. gariepinus occurred in winter season (22%). This was relevant to the study of Feist et al. [13] who verified that **Tetracapsuloides** bryosalmonae used skin and gills for entry then it disseminate to the kidneys through the lymphatic tissues or blood.

Compared to the prevalence of PKD in *O. niloticus*, the prevalence in *C. gariepinus* was lower in all seasons. The general prevalence of PKD in *C. gariepinus* (46.6 %) was lower than *O. niloticus* (83.5 %). The exact cause for this observation is not well-understood. It is well-known that the PKD infection rate is dependent on the host immune system [15]. Also, previous studies revealed that the total leucocytic count is higher in *C.* gariepinus (24.05 \pm 0.050) [3] than in *O.* niloticus (15.0 \pm 4.5) [23]. This could partially explain the lower prevalence of the PKD in *C. gariepinus* than *O. niloticus*. This observation may require further investigations.

Regarding the clinical signs, some affected fishes showed distended abdomen with dark color. exophthalmos, appeared anaemic with pale gills and emaciation. These findings agreed with those reported by others [10, 11, 19 25]. The anaemia observed with PKD could be attributed to the loss of haemopoeitic tissues in the kidney and spleen caused by the Myxozoan parasites. Fernandez-de-Luco et al. [14] reported that abdominal distension and anaemia of rainbow trout affected with PKD were the most obvious clinical signs. Internally, all the affected fishes showed enlargement of the kidneys and spleen and may be ascites but the pathognomonic PM lesion was the enlargement of the posterior kidney that may be protruded ventrally toward the abdominal cavity. These internal findings were in line with those reported by others [9, 18, 12, 26, 29]. In the current study no specific clinical signs were observed in most of the fish although the microscopical, the histopathological PCR results demonstrated the and occurrence of the disease. This was relevant to the results recorded bv Okamura et al. [25] who demonstrated that the identification of the disease may be hindered by absence of clinical signs and hence there was a great possibility for misdiagnosis. Similar observations were reported by Buck et al. [5] as they were No gross changes observed in fishes affected with PKD, however histological various examination revealed Myxosporean stages in renal tubules.

Microscopically, different types of mature Myxosporean spores were identified by examination of the fresh preparations of affected kidneys. Similar results were recorded by El-Mansy and Abdel-Ghaffar identified They [10]. seventeen Myxosporean species belong to the genera Myxobolus, Tiangula and Chloromyxum in Tilapian fishes from the River Nile at El-Rahawy drain. The presence of different Myxosporean spores together in the kidney of almost all examined Tilapian fishes could be attributed to two reasons, first that mature spores might come from their specific organs to the kidney via blood and in this case the kidney infection could be used as a diagnostic evidence for the presence of Myxosporean parasites and the other reason that some spores may originate as a final stage of the development of PKD cells within the kidney tissues because mature spores were already found together with these stages surrounded with cyst like structures [10]. From this study, it was concluded that PKD occurs with high prevalence in Egypt in both O. niloticus and C. gariepinus. The prevalence was usually higher in O. niloticus than C. gariepinus. This may suggest the urgent need to place a strict control measure because of the great economic losses imposed by the disease since these kinds of fishes represent main source for protein most of Egyptian population.

5. REFERENCES

- 1. Abbass, A.A., Lashein, G.H.A. and Tantawy, A.A. 2006. The prevailing endoparasitic diseases of Catfish "*Clarias gariepinus*" with special reference to the associated pathological changes. *Zag. Vet. J.* **34**: 120-139.
- Abdel-Latif, A.M. 2007. Gill parasitic diseases of some freshwater fishes. PhD. Thesis, Fac. Vet. Med., Benha University.
- 3. Aly, S.M., Abd-Allah, O., Mahmoud, A. and Gafer, H. 2010. Efficiency of Levamisole in Improving the Immune Response of Catfish (*Clarias gariepenus*)

to *Aeromonas hydrophila* Vaccine: Clinico- Pathological Studies. *MAJ* **1**: 8-17.

- Anderson C., Canning E.U. and Okamura B. 1999. 18S rDNA sequences indicate that PKX organism parasitizes Bryozoa. Bulletin of the European Association of Fish Pathologists. *Bull Eur Assoc Fish Pat* 19: 94–97.
- 5. Buck D., Feist S.W. and Clifton-Hadley R.S. 1991. The occurrence of proliferative kidney disease (PKD) in cultured and wild fish: further investigations. *J. Fish Dis.* **14**: 583-588.
- Canning, E.U., Curry, A., Feist, S.W., Longshaw, M. and Okamura, B. 1999. *Tetracapsula bryosalmonae* n. sp. for PKX organism, the cause of PKD in salmonid fish. *Bull Eur Assoc Fish Pat* 19: 203–206
- Canning, E. U., Curry, A., Feist, S. W., Longshaw, M. and Okamura, B. 2000. A new class and order of myxozoans to accommodate parasites of bryozoans with ultrastructural observations on *Tetracapsula bryosalmonae* (PKX organism). J. Euk. Microbiol. 47: 456–468.
- 8. Chilmonczyk, S., Monge, D. and De-Kinkelin, P. 2002. Proliferative kidney disease: cellular aspects of the rainbow trout, *Oncorhynchus mykiss* (Walbaum), and response to parasitic infection. *J Fish Dis* **25**: 217–226
- Eissa, A.E., Abu Mourad, I.M.K. and Borhan T. 2006. A contribution on myxosoma Infection in Cultured *Oreochromis niloticus* in Lower Egypt. *Nature and Science* 4: 40-46.
- El-Mansy, A. and Abdel-Ghaffar, F. 2003. Tilapian proliferative kidney disease (TPKD) and a diagnostic evidence for the presence of myxosporean parasites. J. Egypt. Ger. Soc. Zool. 40D: 139-159.
- El-Matbouli, M. and Hoffman, R. W. 2002. Influence of water quality on the outbreak of proliferative kidney disease: field studies and exposure experiments. *J. Fish Dis.* 25: 459–467.
- Feist, S.W. and Longshaw, M. 2006. The Phylum Myxozoa. In: Fish Diseases and Disorders vol. 1. (Ed P.T.K. Woo), CABI. Publishing, Wallingford, UK. Pp.230-296.
- Feist, S. W., Peeler, E. J., Gardiner, R., Smith, E. and Longshaw, M. 2002. Proliferative kidney disease and renal myxosporidiosis in juvenile salmonids

from rivers in England and Wales. J. Fish Dis. 25: 451–458.

- Fernandez-de-Leuco, D., Peribanez, M.A., Garcial and Cstillo, J.A. 1997.
 Granulomatous myositis in rainbow trout oncorhynchus mykiss affected by proliferative kidney disease (PKD). Dis Aquat Org 31: 49-54.
- Gay, M., Okamura, B. and De- Kinkelin, P. 2001). Evidence that infectious stages of *Tetracapsula bryosalmonae* for rainbow trout *Oncorhynchus mykiss* are present throughout the year. *Dis. Aquat. Org.* 46: 31–40.
- Hedrick, R.P., MacConnell, E. and de-Kinkelin, P. 1993. Proliferative kidney disease of salmonid fish. In: Annual Review of Fish Diseases 3 (ed. by M. Faisal and F. M. Hetrick). Elsevier Sciences, Oxford. Pp.277–290.
- 17. Kent, M.L. and Hedrick, R.P. 1986. Development of the PKX myxosporean in rainbow trout *salmo gairdneri*. *Dis. Aquat. Org.* **1**: 169-182.
- Kent, M.L., Khattra, J., Hedrick, R.P.and Devlin, R.H. 2000. *Tetracapsula renicola* n. sp. (Myxozoa: Saccosporidae), the PKX myxozoan-the cause of proliferative kidney disease of salmonid fishes. *J Parasitol* 86:103–111
- 19. Kent, M.L., Margolis, L. and Corliss, J.O. 1994. The demise of a class of protists: taxonomic and nomenclatural revisions proposed for the protist phylum Myxozoa Grasse, 1970. *Can J Zool* **72**: 932–937
- Longshaw, M., Le Deuff, R.M., Harris, A.F. and Feist, S.W. 2002. Development of proliferative kidney disease in rainbow trout, *Oncorhynchus mykiss* (Walbaum), following short-term exposure to *Tetracapsula bryosalmonae* infected bryozoans. J. Fish Dis. 25: 443–449.
- 21. Lucky, Z. 1977. Methods for diagnosis of fish diseases AMERIND publication company, BVT. LTd. New Helhi, Bombay, Calcutta and New York.
- 22. MacConnell, E. and Peterson, J. E. 1992. Proliferative kidney diseasein feral cutthroat trout from a remote Montana reservoir: a first case. J. Aquat. Anim. Hlth 4, 182–187.
- Martins, M. L., Vieira, F.N., Jerônimo, G.T., Mouriño, J.L.P., Dotta, G., Speck, G.M., Bezerra, A. J.M., Pedrotti, F. S.,

Buglione-Neto, C.C. and Pereira, G. 2009. Leukocyte response and phagocytic activity in Nile tilapia experimentally infected with Enterococcus species. *Fish Physiol Biochem* **35**: 219–222.

- 24. Noga, E.J. 1996. Fish diseases Diagnosis and Treatment. Mosby. New York. Pp.367.
- 25. Okamura, B., Hartikainen, H., schmidtposthaus, H.and Wahli, T. 2011. Life cycle complexity, environmental change and the emerging status of salmonid proliferative kidney disease. *Freshwater Biol.* **56**: 735-753.
- Saulnier, D., Philippe, H. and De Kinkelin, P. 1999. Molecular evidence that the proliferative kidney disease organism unknown (PKX) is a myxosporean. *Dis. Aquat. Org.* 36: 209–212.
- 27. Schmidt-Posthaus, H., Bettge, K., Forster, U., Segner, H.and Wahli, T. 2012. Kidney pathology and parasite intensity in rainbow trout *Oncorhynchus mykiss* surviving proliferative kidney disease: time course and influence of temperature. *Dis Aquat Org* **97**: 207-218.
- Shehab El-Din, M.T. 2008. Studies on some nodular parasitic diseases of fishes. PhD Thesis, Fac. Vet. Med., Benha Univ, Egypt.
- 29. Sterud, E., Forseth, T., Ugedal, O., Poppe, T.T., Jorgensen, A., Bruheim, T., Fjeldstad, H.P. and Mo T.A. 2007. Severe mortality in wild Atlantic salmon salmo salar due to proliferative kidney disease (PKD) caused by *Tetracapsuloid bryosalmonae* (Myxozoa). *Dis Aquat Org* **77**: 191-198.
- 30. Tops, S., Hartikainen, H.L.and Okamura, B. 2009. The effects of infection by *Tetracapsuloides bryosalmonae* (Myxozoa) and temperature on *Fredericella sultana* (Bryozoa). *Int J Parasitol.* **39**: 1003-1010.
- Tops, S. and Okamura, B. 2003. Infection of bryozoans by *Tetracapsuloids bryosalmonae* at sites endemic for salmonid proliferative kidney disease. *Dis Aquat Org* 57: 221-226.
- Wahli, T., Knuesel, R., Bernet, D., Segner, H., Pugovkin, D., Burkhardt-Holm, P., Escher, M. and Schmidt-Posthaus, H. 2002. Proliferative kidney disease in Switzerland: current state of knowledge. *J. Fish Dis.* 25: 491–500.



دراسات وبائية عن مرض الكلى التكاثرى فى أسماك البلطى النيلى وأسماك القط الإفريقى إيمان ابراهيم محمد سرور¹، كريمة فتحى محروس²، اسماعيل عبدالمنعم عيسى¹، أمانى عبدالرحمن عباس ¹قسم أمراض الأسماك ورعايتها – كلية الطب البيطرى – جامعة بنها، ²قسم بيولوجيا الخلية – المركز القومى للبحوث، ³قسم أمراض الأسماك ورعايتها – كلية الطب البيطرى – جامعة قناة السويس

الملخص العربى

مرض الكلى التكاثرى من الامراض التى تسبب خسائر اقتصادية كبيرة في المزارع السمكية في جميع أنحاء العالم. وتهدف هذه الدراسة لتحديد معدل الإصابة ومسببات أمراض الكلى التكاثري (PKD) في البلطي النيلي واسماك القط الإفريقي التى تم الحصول عليها من الرياح التوفيقي وروافده. لتحقيق هذا الهدف، تم تجميع عدد 500 من الأسماك منها 266 من البلطي و 244 من اسماك القط الإفريقى. وتم تشريح الأسماك و فحص الكلى ظاهريا وملاحظة أى تغيرات شكلية او عقد مرضيه .تم الفحص المباشر للعينات الكلوية تحت الميكروسكوب وايضا تم الفحص بعد صباغة الشرائح بصبغة جيمسا. أظهرت النتائج حدوث المرض في البلطي النيلي وكانت أعلى معدلات الاصابة في فصل الخريف (85.08%) والربيع (91.94 %) واقل فى فصل الصيف (76.00%) والشتاء (73.53%). وكانت نسبة حدوث وقوع المرض في أسماك القط عالية في فصل الشتاء (0.67%) ومنخفض في فصل الربيع معدادية غير محدد وهى عبارة عن في أسماك القط عالية في فصل الشتاء (0.70%) ومنخفض في فصل الربيع المرضية غير محدد وهى عبارة على في البلطي النيلي ما 83.46 % عن أسماك القط 18.58%. وكانت معظم العلامات المرضية غير محدد وهى عبارة على في البلطي النيلي ما 83.46 % عن أسماك القط عالية عد المرضية غير محدد وهى عبارة على في البلطي النيلي ما 83.46 % عن أسماك القط عالي في المرابيا في معدلات الموساك الديها عقد المرضية غير محدد وهى عبارة على في البلطي النيلي ما 83.46 % عن أسماك القط 18.58%. وكانت معظم العلامات المرضية غير محدد وهى عبارة على ألمان الماكن، وفقر الدم وشحابة الخياشيم مع اليزال بعض الأسماك لديها عقد في العين حول القزحية تظهر وكانها حلقة، كان هناك تضخم في الكلى مع ظهور بعض العقد الكلوية من 0.20–0.60 مم .بناء على المرضية غير محدد وهى عبارة على الماسبب المرض مثل أنواع مختلفة من ميكسوسبورين في البلطي النيلي واسماك النها المرضية من حول القرحية تظهر وكانها لماله المالك مع ظهور بعض العقد الكلوية من 2.00–0.60 مم .بناء على المرضية أسماك القط في الظاهرى تم التقريقة، منهمال المنولي من الخول الأمي القط في المحل منزل أنواع مختلفة من ميكسوسبورين في البلطي النيلي واسماك القط الأمري القل الشكل الظاهرى تم التعرف على مرض الكلى التكاثرى فى هذه الإسماك القط المراك القط المولي الماك الفل المحل مرفر الكل المحل مرض الكلى الماه المرام من الأمرا محل المراك الفل مراكساك الفلو من المماك ا

(مجلة بنها للعلوم الطبية البيطرية: عدد 23 (1)، يونيو 2012: 158-158)